

New Fluorenocrownphanes with Naphthalene Fragments: Synthesis, Structure, Properties, and Interaction with Paraquat

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New crownphanes containing fragments of 1,5-dioxynaphthalene and 2,7-dioxyfluorene or 2,7-dioxyfluorene were synthesized by reduction of the corresponding fluorenonocrownphanes. H₂-Pd/C or Wolff-Kishner methods were utilized. Molecular structure of bis(fluoreno)crownophane with tetraethyleneglycol linkers was confirmed by single-crystal X-ray diffraction. The formation of pseudorotaxane-type host-guest complexes of the obtained crownphanes with paraquat dication was detected by FABMS, ¹H NMR and UV-Vis spectroscopy.

Keywords: Fluorenocrownphanes, fluorenonocrownphanes, reduction, paraquat, pseudorotaxanes.

Новые флуоренокраунофаны с фрагментами нафталина: синтез, структура, свойства и взаимодействие с паракватом

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Восстановлением соответствующих флуоренокраунофанов в системе H₂-Pd/C либо в условиях реакции Кижнера-Вольфа получены новые флуоренокраунофаны, содержащие два фрагмента 2,7-диоксифлуорена либо фрагменты 2,7-диоксифлуорена и 1,5-диоксинафталина. Строение молекулы бис(флуорено)краунофана с фрагментами тетраэтиленгликоля было подтверждено методом РСА. Методами ББА-масс-спектрометрии, ¹H ЯМР и электронной спектроскопии установлено образование псевдоротаКСанов при взаимодействии полученных краунофанов с паракват-дикатионом.

Ключевые слова: Флуоренокраунофаны, флуоренонокраунофаны, восстановление, паракват, псевдоротаКСаны.

Introduction

The contemporary chemistry of cyclophanes has shifted its focus from pure synthetic and structural studies to the study of properties which give way to practical applications. In the last decade cyclophanes and crownphanes – their polyether analogs, have been actively employed for the synthesis of supramolecular structures such as pseudorotaxanes, rotaxanes and catenanes.^[1–3] The practical goal of these studies was to create novel compounds that are able to form strong complexes with electron-deficient substrates which can then be used to construct supramolecular and sensor systems.^[4–7]

The fluorene moiety, as well as its derivative fluorenone, has a large π -electron system and the removal of the carbonyl group results in increased electron density and the uniformity of its distribution. The methylene protons in position 9 can act as a “probe” as they are situated out of the plane of the aromatic rings and should be most sensitive to the spatial environment changes of the molecule, which can be used for study of complexation properties by NMR. Moreover, the fluorene fragment has excellent fluorescent and electrochemical properties and cyclophanes constructed from fluorene play an important role in the creation of organic light emitting diodes (OLED),^[8,9] charge transporters,^[10,11] transistors,^[12] sensors,^[13] and two-photon absorption materials.^[14] Though a large number of compounds has been described, the search for novel macrocyclic compounds including the fluorene moiety is still actual.

We have earlier described the synthesis of fluoreno-crownphanes containing 2,7-dioxyfluorene and hydroquinone or 4,4'-dioxybiphenyl, the qualitative assessment of their paraquat complexes demonstrated the perspectives of further investigation of their complexation properties.^[15] In the light of the above, the current research was aimed at describing the influence of the second aromatic ring on the properties of fluorenocrownphanes. To achieve this we have synthesized novel fluorenocrownphanes containing either two fluorene or one fluorene and one naphthalene fragments and performed a qualitative study of the complex formation with model electron-deficient substrate – paraquat-dication in order to evaluate the perspectives of the further studies.

Experimental

The ¹H and ¹³C NMR spectra were recorded on Varian VXR-300 (¹H) and Bruker Avance DRX 500 (¹³C) instruments with the operating frequencies of 300 and 125.76 MHz correspondingly, the ¹H spectra of the complexes were recorded in a mixture of CD₃CN–CDCl₃ (4:3, v/v). The EI mass spectra were obtained on a MX-1321 mass spectrometer (70 eV, direct injection), the FAB mass spectra were obtained on a VG 7070EQ mass spectrometer (Xe, 8 kV) in a matrix of 3-nitrobenzyl alcohol. The UV-Vis spectra were recorded on a Specord M-40 spectrophotometer. The elemental analysis was conducted on a EuroVector CHNS analyzer. The melting points were measured using open capillaries and were not corrected. The purity of all the compounds synthesized were monitored by TLC (Al₂O₃/Al plates by Fluka and SiO₂ Sorbfil UV-254). Preparative flash chromatography was performed on silica gel 60 (<63 μ m, Merck KGaA). X-Ray structure study of compound **2c** was performed on an «Xcalibur-3» diffractometer (MoK α

radiation, CCD-detector, graphite monochromator, ω -scan, $2\theta_{\max} = 60^\circ$). The structure was solved by direct methods using SHELXTL software.^[16] The positions of hydrogen atoms were derived from differential electron density synthesis and refined within riding model with fixed isotropic displacement parameters with $U_{\text{iso}} = 1.2U_{\text{eq}}$ of a non-hydrogen atom bound to the current hydrogen atom. The structure was refined by full-matrix least squares technique on F^2 with anisotropic displacement parameters for non-hydrogen atoms with $wR_2 = 0.054$ for 10235 reflections ($R_1 = 0.039$ for 5510 reflections with $F > 4\sigma(F)$, $S = 0.963$), CCDC 1410263. 1,5-Dimethoxynaphthalene was purchased from commercial sources. Fluorenocrownphanes **1a-c,e-g**,^[17–19] and paraquat bis(hexafluorophosphate) **6-2PF₆**^[20] were obtained by previously described procedures.

Synthesis

8, 11, 14, 17, 20, 23, 35, 38, 41, 44, 47, 50-Dodecaoxaheptacyclo[49.3.1.1^{3,7}.1^{24,28}.1^{30,34}.0^{4,54}.0^{27,31}]octapentaconta-1(55),3(58),4,6,24(57),25,27,30(56),31,33,51,53-dodecaen-2,29-dione (1d). To a suspension of 5.52 g (0.04 mol) of anhydrous potassium carbonate in 600 mL of dry DMF was added with stirring a solution of 0.01 mol of 2,7-dihydroxyfluorenone and 0.01 mol of the corresponding bistosylate^[17] in 400 mL of dry DMF. The addition was performed for 10 h while the temperature of the reaction mixture was maintained at 80 °C. When the addition was finished the mixture was stirred for extra 40 h at the same temperature. The cooled reaction mixture was filtered and the solvent evaporated *in vacuo*. The residue was extracted with two toluene portions of 600 and 400 mL, the solvent was evaporated *in vacuo* and the residue was subjected to chromatographic purification (SiO₂, CHCl₃–CH₃OH, 100:3) and recrystallization from toluene. Orange powder. Yield: 4.39 g (53 %), m.p. 172–173 °C. Found, %: C 66.87, H 6.40. C₄₆H₅₂O₁₄. Calculated, %: C 66.65, H 6.32. *m/z* (EI) I_{rel}^+ , %: 828 [M]⁺ (24), 239 (15), 211 (12), 45 (100). UV-Vis (C₄H₈O₂) λ_{max} (lg ϵ) nm: 263 (5.04), 271 (5.03), 302 (4.09), 314 (4.06), 468 (2.66). ¹H NMR (CDCl₃) δ_{H} ppm (*J*, Hz): 3.67–3.77 (m, 24H, CH₂O), 3.87 (t, 8H, *J* 4.5, CH₂O), 4.04 (t, 8H, *J* 4.5, CH₂O), 6.77 (dd, 4H, *J* 8.4, *J* 2.2, H_a), 6.91 (d, 4H, *J* 2.2, H_b), 7.00 (d, 4H, *J* 8.4, H_c). ¹³C NMR (CDCl₃) δ_{C} ppm: 67.8, 69.6, 70.7, 70.9 (2C), 109.9, 120.3, 120.5, 135.6, 137.3, 158.9, 193.2.

Typical procedure for the synthesis of fluorene crownphanes 2a-d. A stream of hydrogen was passed for 0.5 h through a 10 % Pd/C suspension (0.1 g) being stirred in a mixture of CH₃OH–PhCH₃ (3:1, 40 mL) maintained at 55–60 °C, then a solution (0.5 mmol) of the corresponding fluorenone crownophane **1a-d** with the same temperature and in the same solvent mixture (80 mL) was added. The stirring was continued, while maintaining the stream of hydrogen and the set temperature until full conversion of the starting material was achieved (20–25 h, TLC). The hot reaction mixture was filtered, the precipitate washed with hot toluene (5×20 mL), the combined filtrates evaporated *in vacuo* and the residue was recrystallized from ethanol.

8, 11, 14, 26, 29, 32-Hexaoxaheptacyclo[31.3.1.1^{3,7}.1^{15,19}.1^{21,25}.0^{4,36}.0^{18,22}]tetraconta-1(37),3(40),4,6,15(39),16,18,21(38),22,24,33,35-dodecaene (2a). Colorless crystals. Yield: 247 mg (92 %), m.p.: 241–242 °C. Found, %: C 76.29, H 6.11. C₃₄H₃₂O₆. Calculated, %: C 76.10, H 6.01. *m/z* (EI) I_{rel}^+ , %: 536 [M]⁺ (100), 268 (18), 197 (15). UV-Vis (C₄H₈O₂) λ_{max} (lg ϵ) nm: 274 (4.58), 313 (3.93). ¹H NMR (DMSO-*d*₆) δ_{H} ppm (*J*, Hz): 2.98 (s, 4H, H_a), 3.75–3.82 (m, 8H, CH₂O), 4.06–4.13 (m, 8H, CH₂O), 6.64 (br.s., 4H, H_a), 6.86 (dd, 4H, *J* 8.1, 1.6, H_b), 7.54 (d, 4H, *J* 8.4, H_c).

8, 11, 14, 17, 29, 32, 35, 38-Octaoxaheptacyclo[37.3.1.1^{3,7}.1^{18,22}.1^{24,28}.0^{4,42}.0^{21,25}]hexatetraconta-1(43),3(46),4,6,18(45),19,21,24(44),25,27,39,41-dodecaene (2b). Colorless crystals (ethanol). Yield: 193 mg (62 %), m.p.: 148 °C. Found, %: C 73.11, H 6.62. C₃₈H₄₀O₈. Calculated, %: C 73.06, H 6.45. *m/z* (EI) I_{rel}^+ , %: 624

[M]⁺ (100), 312 (14), 198 (16). UV-Vis (CH₃CN) λ_{max} (lgε) nm: 274 (4.26), 315 (3.58). ¹H NMR (CDCl₃) δ_H ppm (*J*, Hz): 3.29 (s, 4H, H₉), 3.78 (s, 8H, CH₂O), 3.89 (t, 8H, *J* 4.4, CH₂O), 4.04 (t, 8H, *J* 4.4, CH₂O), 6.76 (s, 4H, H_a), 6.83 (d, 4H, *J* 8.4, H_b), 7.35 (d, 4H, *J* 8.1, H_c). ¹³C NMR (CDCl₃) δ_C ppm: 36.9, 68.1, 70.1, 71.3, 111.4, 113.8, 119.4, 134.8, 144.4, 157.7.

8,11,14,17,20,32,35,38,41,44-Decaoxaheptacyclo[43.3.1.1^{3,7}.1^{21,25}.1^{27,31}.0^{4,48}.0^{24,28}]dopentaconta-1(49),3(52),4,6,21(51),22,24,27(50),28,30,45,47-dodecaene (2c). Colorless crystals. Yield: 189 mg (53 %), m.p.: 138 °C. Found, %: C 70.53, H 6.69. C₄₂H₄₈O₁₀. Calculated, %: C 70.77, H 6.79. *m/z* (EI) *I*_{rel.}⁺, %: 712 [M]⁺ (100), 355 (8), 198 (16). UV-Vis (CH₃CN) λ_{max} (lgε) nm: 273 (4.32), 315 (3.64). ¹H NMR (CDCl₃) δ_H ppm (*J*, Hz): 3.43 (s, 4H, H₉), 3.72–3.79 (m, 16H, CH₂O), 3.90 (t, 8H, *J* 4.4, CH₂O), 4.02 (t, 8H, *J* 4.4, CH₂O), 6.78–6.83 (m, 8H, H_a, H_b), 7.34 (d, 4H, *J* 8.1, H_c). ¹³C NMR (CDCl₃) δ_C ppm: 36.8, 67.8, 69.8, 70.7, 70.9, 111.0, 113.5, 119.5, 134.7, 144.4, 157.6.

8,11,14,17,20,23,35,38,41,44,47,50-Dodecaoxaheptacyclo[49.3.1.1^{3,7}.1^{24,28}.1^{30,34}.0^{4,54}.0^{27,31}]octapentaconta-1(55),3(58),4,6,24(57),25,27,30(56),31,33,51,53-dodecaene (2d). Colorless crystals. Yield: 128 mg (32 %), m.p.: 152 °C. Found, %: C 69.07, H 7.09. C₄₆H₅₆O₁₂. Calculated, %: C 68.98, H 7.05. *m/z* (EI) *I*_{rel.}⁺, %: 800 [M]⁺ (92), 399 (5), 198 (100). UV-Vis (CH₃CN) λ_{max} (lgε) nm: 275 (4.55), 316 (3.89). ¹H NMR (CDCl₃) δ_H ppm (*J*, Hz): 3.56 (s, 4H, H₉), 3.68–3.75 (m, 24H, CH₂O), 3.85 (t, 8H, *J* 4.9, CH₂O), 4.05 (t, 8H, *J* 4.8, CH₂O), 6.80 (dd, 4H, *J* 8.4, 1.6, H_b), 6.89 (s, 4H, H_a), 7.38 (d, 4H, *J* 8.3, H_c). ¹³C NMR (CDCl₃) δ_C ppm: 36.9, 67.7, 69.8, 70.7, 70.8, 70.9, 111.2, 113.4, 119.5, 134.7, 144.4, 157.6.

Typical procedure for the synthesis of fluorene crownphanes 2e-g. Hydrazine hydrate (0.8 mL, 16.4 mmol) was added to a suspension of 0.4 mmol of the corresponding crownophane **1e-g** in 8 mL of diethylene glycol under an inert atmosphere and the mixture was stirred at 130 °C until a homogenous bright yellow solution was formed. The mixture was cooled to 100 °C and 0.2 mL of KOH solution was added (1 g KOH in 5.6 mL of diethylene glycol) and the temperature was raised up to 200 °C. This temperature was maintained for 5 min, then cooled to 100 °C and 12 mL of acidified water were added. The cooled mixture was filtered, the residue was washed several times with water and dried. Column chromatography on SiO₂ was then performed using CH₂Cl₂–2-propanol 100:1 as the eluent, followed by recrystallization from a suitable solvent.

8,11,14,17,28,31,34,37-Octaoxaheptacyclo[36.3.1.1^{3,7}.0^{4,41}.0^{18,23}.0^{22,27}]tritetraconta-1(42),3(43),4,6,18,20,22,24,26,38,40-undecaene (2e). Recrystallized from 1-butanol. Colorless crystals. Yield: 202 mg (86 %), m.p.: 138–138.5 °C. Found, %: C 71.42, H 6.70. C₃₅H₃₈O₈. Calculated, %: C 71.65, H 6.53. *m/z* (EI) *I*_{rel.}⁺, %: 586 [M]⁺ (100), 293 (32), 198 (14), 45 (21). UV-Vis (CH₃CN) λ_{max} (lgε) nm: 226 (4.72), 281 (4.39), 313 (3.98), 326 (3.93). ¹H NMR (CDCl₃) δ_H ppm (*J*, Hz): 3.07 (s, 2H, H₉), 3.69–4.00 (m, 16H, CH₂O), 4.04–4.22 (m, 8H, CH₂O), 6.56 (d, 2H, *J* 7.5, H_{2,6}), 6.77 (s, 2H, H_a), 6.84 (d, *J* 8.1, 2H, H_b), 7.04 (t, 2H, *J* 7.9, H_{3,7}), 7.35 (d, 2H, *J* 8.4, H_c), 7.74 (d, 2H, *J* 8.4, H_{4,8}). ¹³C NMR (CDCl₃) δ_C ppm: 36.6,

67.5, 68.4, 69.8, 70.0, 71.0, 71.1, 77.0, 105.2, 111.9, 114.0, 114.2, 119.1, 124.8, 126.4, 134.7, 144.4, 153.9, 157.3.

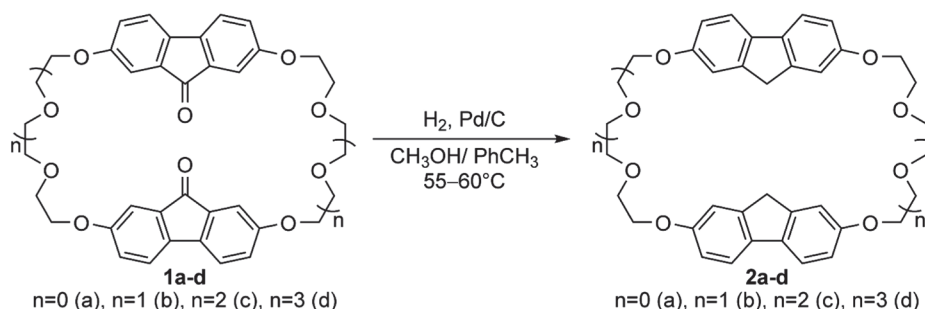
8,11,14,17,20,31,34,37,40,43-Decaoxaheptacyclo[42.3.1.1^{3,7}.0^{4,47}.0^{21,26}.0^{25,30}]nonatetraconta-1(48),3(49),4,6,21,23,25,27,29,46-decaene (2f). Recrystallized from 2-propanol. Colorless crystals. Yield: 182 mg (66 %), m.p.: 137.5–138 °C. Found, %: C 69.70, H 7.03. C₃₉H₄₆O₁₀. Calculated, %: C 69.42, H 6.87. *m/z* (EI) *I*_{rel.}⁺, %: 674 [M]⁺ (100), 337 (5), 198 (7), 45 (18). UV-Vis (CH₃CN) λ_{max} (lgε) nm: 226 (4.72), 280 (4.53), 313 (4.12), 327 (4.06). ¹H NMR (CDCl₃) δ_H ppm (*J*, Hz): 3.45 (s, 2H, H₉), 3.63–3.82 (m, 16H, CH₂O), 3.83–3.97 (m, 8H, CH₂O), 3.98–4.17 (m, 8H, CH₂O), 6.53 (d, 2H, *J* 7.8, H_{2,6}), 6.81–6.93 (m, 4H, H_a, H_b), 7.09 (t, 2H, *J* 7.9, H_{3,7}), 7.43 (d, 2H, *J* 8.1, H_c), 7.77 (d, 2H, *J* 8.4, H_{4,8}). ¹³C NMR (CDCl₃) δ_C ppm: 36.8, 67.7, 67.9, 69.7, 69.8, 70.7, 70.8, 70.9 (2C), 77.0, 105.3, 111.5, 113.4, 114.3, 119.4, 124.9, 126.5, 134.7, 144.3, 154.0, 157.4.

8,11,14,17,20,23,34,37,40,43,46,49-Dodecaoxaheptacyclo[48.3.1.1^{3,7}.0^{4,53}.0^{24,29}.0^{28,33}]pentapentaconta-1(54),3(55),4,6,24,26,28,30,32,50,52-undecaene (2g). Recrystallized from methanol. Colorless crystals. Yield: 268 mg (88 %), m.p.: 102–102.5 °C. Found, %: C 67.82, H 7.10. C₄₃H₅₄O₁₂. Calculated, %: C 67.70, H 7.13. *m/z* (EI) *I*_{rel.}⁺, %: 762 [M]⁺ (100), 307 (29), 198 (23). UV-Vis (CH₃CN) λ_{max} (lgε) nm: 226 (4.85), 280 (4.53), 313 (4.11), 326 (4.06). ¹H NMR (CDCl₃) δ_H ppm (*J*, Hz): 3.61 (s, 2H, H₉), 3.63–3.77 (m, 24H, CH₂O), 3.80–3.93 (m, 8H, CH₂O), 4.05–4.16 (m, 8H, CH₂O), 6.64 (d, 2H, *J* 7.5, H_{2,6}), 6.85 (dd, 2H, *J* 8.4, 1.9, H_b), 6.94 (br.s., 2H, H_a), 7.23 (t, 2H, *J* 8.1, H_{3,7}), 7.45 (d, 2H, *J* 8.1, H_c), 7.78 (d, 2H, *J* 8.4, H_{4,8}). ¹³C NMR (CDCl₃) δ_C ppm: 36.9, 67.7, 67.8, 69.7, 69.8, 70.7 (2C), 70.8 (2C), 70.9, 71.0, 77.0, 105.4, 111.4, 113.5, 114.4, 119.5, 125.0, 134.7, 144.3, 154.1, 157.5.

2,7-Dimethoxy-9H-fluorene (4). A mixture of 2.4 g (0.01 mol) of 2,7-dimethoxyfluorenone (3)^[15] and 7.3 mL (0.15 mol) of hydrazine hydrate in 40 mL of ethylene glycol was heated in an inert atmosphere at 130–140 °C until a homogenous solution was formed. The mixture was cooled to 100 °C and 1.7 g (0.03 mol) of KOH was added, followed by heating to 190 °C and maintaining the temperature for 5 min. Then an equal volume of water was added to a cooled to 95 °C mixture and the formed precipitate was filtered, washed with water, dried and recrystallized from toluene with the addition of some Al₂O₃. Colorless crystals. Yield: 1.9 g (86 %), m.p. 173 °C. Found, %: C 79.60, H 6.41. C₁₅H₁₄O₂. Calculated, %: C 79.62, H 6.24. *m/z* (EI) *I*_{rel.}⁺, %: 226 [M]⁺ (100), 211 (89). ¹H NMR (CDCl₃) δ_H ppm (*J*, Hz): 3.81 (s, 2H, H₉), 3.84 (s, 6H, CH₃), 6.89 (dd, 2H, *J* 8.4, 2.2, H_b), 7.06 (d, 2H, *J* 2.2, H_a), 7.56 (d, 2H, *J* 8.4, H_c).

Results and Discussion

Bis(fluoreno)crownphanes **2a-d** were synthesized in a manner analogous to described in our previous paper^[15] by reduction of the corresponding fluorenone crownphanes **1a-d** with hydrogen in the presence of 10 % Pd/C in a 3:1 mixture of methanol and toluene (Scheme 1). The reaction



Scheme 1.

progress was monitored by TLC. After the proper treatment of the reaction mixture and recrystallization of the target compounds from ethanol they were isolated as colorless crystals with 52–92 % yields.

However the attempt to reduce crownphanes **1e-g** with a naphthalene fragment under the same conditions did not give satisfactory results. At first, the 9-hydroxy derivative (identified by mass spectrometry) was rapidly accumulated in the reaction mixture. Its further reduction, though, proceeded very slowly compared to the first reaction step, which resulted in the formation of a complex mixture of highly polar compounds instead of the target compound (according to TLC). Changing various reaction parameters, such as increasing the temperature to 60 °C, addition of toluene in order to increase the solubility of the starting crownophane or addition of acetic acid as a catalyst, did not lead to satisfactory results.

Two other general methods for the reduction of a ketone group to a methylene one are the Clemmensen and Wolff–Kishner reactions. The first method could not be utilized due to the low solubility of the starting compounds in water, whilst the reduction of the model 2,7-dimethoxyfluorenone **3** with hydrazine hydrate in ethylene glycol in the presence of KOH afforded 86 % of 2,7-dimethoxyfluorene **4**.

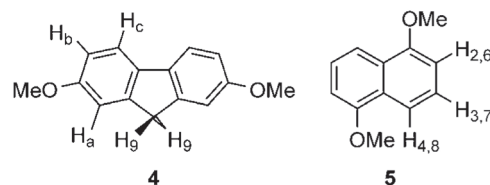
Reduction of **1e-g** under the above conditions was found possible only for the crownophane **1g**, which has the highest solubility. However, when diethylene glycol was used as the solvent and a large excess (40 eq) of hydrazine hydrate was added, all crownphanes **1e-g** were successfully reduced. The excess of hydrazine hydrate and the absence of acidic catalysts prevent the formation of azine side products. After the treatment of the reaction mixture, chromatographic separation and recrystallization the target crownphanes **2e-g** were isolated with 66–88 % yields (Scheme 2).

The structure of the synthesized crownphanes **2a-g** was confirmed by NMR spectroscopy, mass spectrometry, elemental analysis, and by X-ray diffraction data for compound **2c**.

A typical feature of mass spectra (electron impact) of all the synthesized fluorenocrownphanes **2a-g** is the presence of doubly charged molecular ions M^{2+} , which correspond to half the molecular weight mass, as well as fragmentary ions with the m/z ratios equal to 198 (fluorene residue) and 45 (side chain fragments).

A signal set typical for ethylene glycol oligomers is observed in the 1H NMR spectra of fluorenocrownphanes **2a-g** in the δ 4.29–3.55 ppm region. The protons of the

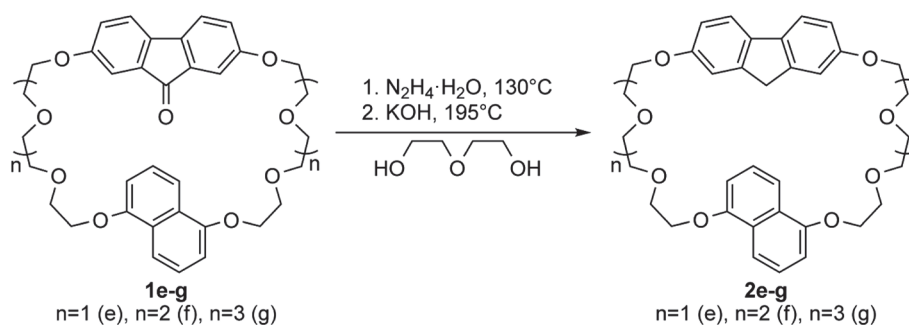
fluorene fragment are presented as a typical set of signals: a broadened singlet (H_a), two doublets (H_b , H_c), and a singlet of H_9 methylene group. The protons of 1,5-dioxynaphthalene are represented by two doublets ($H_{2,6}$ and $H_{4,8}$) and a triplet ($H_{3,7}$). For bis(fluoreno)crownphanes **2a-d** the signals of the aromatic protons undergo an upfield shift compared to their position in the spectrum of 2,7-dimethoxyfluorene **4**. The values of the upfield shift are the greatest for H_a -protons and diminishing simultaneously with increase of the ring size (Table 1).



Similarly, the signals of all aromatic protons with the exception of H_c protons in **2e-g** undergo upfield shifts relative to the position of these signals in the spectra of model compounds **4** and **5**. Such a spectral pattern is a result of mutual shielding of the opposite aromatic fragments of the crownphanes (Table 1). The magnitude of these shifts is decreased with the increase of the ring size. Signals of H_9 protons for the above reasons exhibit the greatest upfield shifts, particularly noticeable for bis(fluoreno)crownphanes **2a-d**. The singlet form of these protons signal is, obviously, caused by the rapid rotation of the aromatic fragments of fluorenocrownphanes **2a-g** on the NMR time scale. The observed spectral pattern indicates that the aromatic fragments of the crownphanes are spatially close in solutions even in the cases of **2d** and **2g** (according to 1H NMR), where they are linked by pentaethylene glycol residues. Obviously, in solutions the stacking interactions take place with their intensity lowered with the increase of the ring size.

Structure the bis(fluoreno)crownophane **2c** in which the fluorene fragments are linked with tetraethylene glycol residues was confirmed by X-ray structure study. The suitable crystals were obtained by crystallization from acetonitrile. Two molecules (A and B) that have differences in some geometric parameters are identified in the independent part of a unit cell. Each molecule is in a particular position relative to the inversion center; as a result half of the molecule is symmetrically independent.

In both molecules the π -systems of the fluorene fragments are almost not overlapped. The conformational flex-



Scheme 2.

Table 1. Absolute (δ , ppm) and relative ($\Delta\delta^1$, ppm) chemical shifts of the aromatic protons² of crownphanes **2a-g** and model compounds **4** and **5** in CDCl₃ at 298 K.

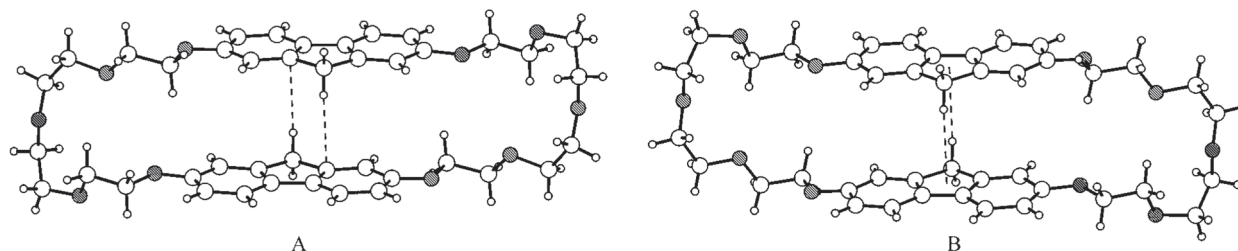
Compound	H ₉		H _a		H _b		H _c		H _{2,6}		H _{3,7}		H _{4,8}	
	δ	$-\Delta\delta$	δ	$-\Delta\delta$	δ	$-\Delta\delta$	δ	$-\Delta\delta$	δ	$-\Delta\delta$	δ	$-\Delta\delta$	δ	$-\Delta\delta$
2a ³	2.98	0.83	6.64	0.42	6.86	0.03	7.54	0.02	–	–	–	–	–	–
2b	3.29	0.52	6.76	0.30	6.83	0.06	7.35	0.21	–	–	–	–	–	–
2c	3.43	0.38	6.81 ⁴	0.25	6.81 ⁴	0.08	7.34	0.22	–	–	–	–	–	–
2d	3.56	0.25	6.89	0.17	6.80	0.09	7.38	0.18	–	–	–	–	–	–
2e	3.07	0.74	6.77	0.29	6.84	0.05	7.35	0.21	6.56	0.27	7.04	0.32	7.74	0.09
2f	3.45	0.36	6.88 ⁴	0.18	6.85 ⁴	0.04	7.43	0.13	6.54	0.29	7.09	0.27	7.76	0.07
2g	3.61	0.20	6.94	0.12	6.85	0.04	7.44	0.12	6.64	0.19	7.23	0.13	7.78	0.05
4	3.81	–	7.06	–	6.89	–	7.56	–	–	–	–	–	–	–
5	–	–	–	–	–	–	–	–	6.83	–	7.36	–	7.83	–

¹ $\Delta\delta$ is equal to the difference of chemical shifts of the corresponding protons in compounds **2a-d** and **4**, and compounds **2e-f** and **4**, **5**, correspondingly.

²The proton enumeration was given above.

³In DMSO-*d*₆.

⁴The signals of H_a and H_b overlap, which hinders the determination of the exact signal positions.

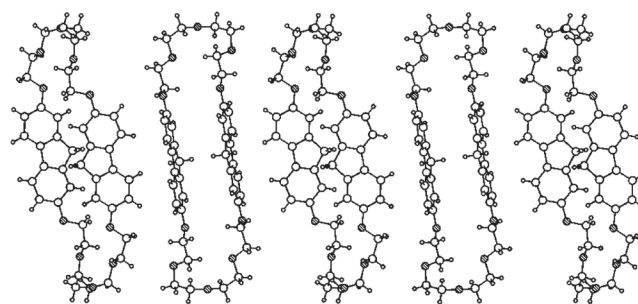
**Figure 1.** Molecules A and B of compound **2c**, identified in the independent part of the unit cell.

ibility of tetraethylene glycol residues provide a shift of fluorene fragments respectively each other in the direction perpendicular to the most elongated dimension of the triple ring. The main difference between A and B molecules is the position of the fluorene fragments with respect to each other along the linker chain (Figure 1): in A the fluorene fragments are slightly shifted compared to B. The mutual disposition of the fluorene fragments is stabilized by two symmetric intramolecular C–H \cdots π hydrogen bonds C₇–H_{7ab} \cdots C₈' (H \cdots C 2.76 Å, C–H \cdots C 145°) for molecule A, and B C₇–H_{7ba} \cdots X (where X is the geometric center of the C₁–C₆–C₇–C₈–C₁₃ ring) (H \cdots X 2.72 Å, C–H \cdots X 153°) for molecule B.

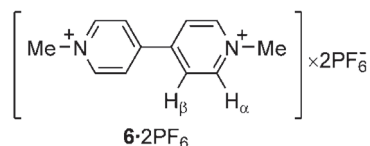
In crystals, molecules A and B interleave forming stacks (Figure 2) lying in the crystallographic plane (1 0 –1). Inside the stacks the molecules are linked with C–H \cdots π intermolecular hydrogen bonds C₁₂–H_{12a} \cdots C_{10b}' (π) (–x, 2–y, –z) H \cdots C 2.86 Å C–H \cdots C 138°; C₂–H_{2aa} \cdots C_{1b}' (π) (–x, 2–y, –z) H \cdots C 2.69 Å C–H \cdots C 152°; C₂–H_{2aa} \cdots C_{6b}' (π) (–x, 2–y, –z) H \cdots C 2.85 Å C–H \cdots C 141°.

The neighboring stacks (Figure 3) are linked with intermolecular hydrogen bonds of C_{15a}–H₁₅ \cdots C_{12b}' (π) (–x, –0.5+y, 0.5–z) H \cdots C 2.76 Å C–H \cdots C 141° and C_{16b}–H \cdots C_{2b}' (π) (x, 2.5–y, –0.5+z) H \cdots C 2.82 Å C–H \cdots C 116°.

Complex of spectral methods was used to demonstrate that all previously synthesized fluorene- and fluorenono-crownphanes form host-guest complexes with paraquat

**Figure 2.** Interleaved molecules A and B in a crystal.

bis(hexafluorophosphate) **6**·2PF₆, which is one of the most studied electron-deficient guests in macrocyclic chemistry. [17–19,21–24] This methodology was used for the studies of the crownphanes **2a-g** as well.



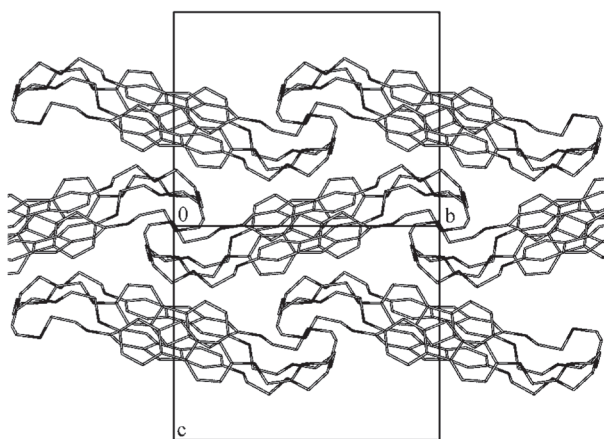


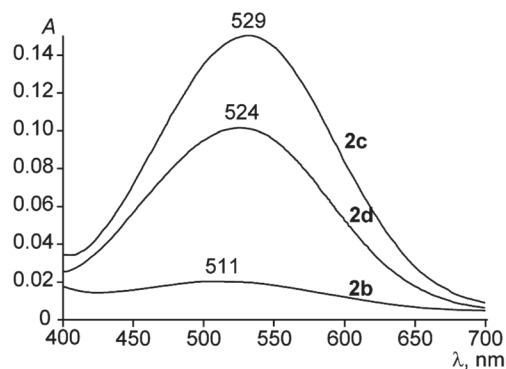
Figure 3. The packing of the stacks in a crystal.

For the qualitative assessment of the possibility of inclusion complexes formation and their relative stability the competitive complex formation method was employed under the FAB mass spectrometry conditions.^[25–27] Two solutions containing equimolar amounts of crownophanes **2b–d** or **2e–g** and 3 equivalents of **6**·2PF₆ in 3-nitrobenzyl alcohol were subjected to mass spectral analysis. In the mass spectra, besides the initial crownophanes peaks, the peaks of ions corresponding to the formation of a crownophane-paraquat 1:1 complex with the loss of a hexafluorophosphate anion were observed (Table 2).

Table 2. Normalized peak intensities of the complex ions of crownophanes **2b–g** and paraquat **6**·2PF₆.

Ion	<i>m/z</i>	<i>I</i> , %
[2b @ 6 ·PF ₆] ⁺	955	–
[2c @ 6 ·PF ₆] ⁺	1043	–
[2d @ 6 ·PF ₆] ⁺	1131	–
[2e @ 6 ·PF ₆] ⁺	917	38
[2f @ 6 ·PF ₆] ⁺	1005	100
[2g @ 6 ·PF ₆] ⁺	1093	50

Such a spectral pattern is typical for most of rotaxanes and pseudorotaxanes^[28–31] and indicates the formation



of pretty stable complexes of fluorencrownophanes **2b–g** with paraquat **6**·2PF₆, which supposedly possess a pseudorotaxane-like structure. Among the fluorencrownophanes **2e–g** with naphthalene fragments the most intensive molecular peak of a complex ion observed for the crownophane **2f** where the aromatic fragments are linked by tetraethylene glycol residues, while less intensive peaks are observed for crownophanes **2g** and **2e**. As the intensity ratio for the peaks of complex ions of structurally similar ligands correlates to the stability of the complexes in the first approximation, it can be suggested that under the mass spectral experiment conditions the most stable complex is formed in the case of crownophane **2f**. In the case of bis(fluoreno)crownophanes **2b–d** the intensity of peaks of the corresponding complex ions was very low, which proves only the fact of formation of these complexes without the possibility to make any conclusions about their stability. The peaks of the ions corresponding to [**2a**@**6**·PF₆]⁺ complex particles were not detected under any experimental conditions. This allows to conclude **2a** does not form a complex with paraquat **6**·2PF₆ under the conditions of a mass spectral experiment.

During the addition of paraquat **6**·2PF₆ to acetonitrile solutions of fluorencrownophanes **2b–g** an intense pink or crimson-red color appears. A broad band appears in the visible region of the UV-Vis spectra (496–530 nm). The appearance of this band is due to the formation of a charge transfer complex (CTC) between the π-donor aromatic cyclophane fragments and the π-acceptor dipyridylic frame of paraquat-dication **6**²⁺ (Figure 4) in the cavity of the macrocycle.^[33–35] It indicates the formation of relatively stable host-guest complexes, similarly to those observed previously.^[15]

The most intensive CTC bands are observed for crownophanes **2c** and **2f** with tetraethylene glycol, which is due to the greater stability of the corresponding complexes. Crownophane **2a** was not studied by this method because of its insufficient solubility. However, even when such solvents as CH₃NO₂ and DMF were used, no visual change of color was observed, which indicates the absence of formation of the corresponding complex particles.

At a qualitative level complex formation can also be studied by ¹H NMR.^[17–19,21–24]

The measurements were made for fixed concentrations of components in a mixture of CD₃CN–CDCl₃ (4:3). For comparison, spectra of paraquat and the corresponding

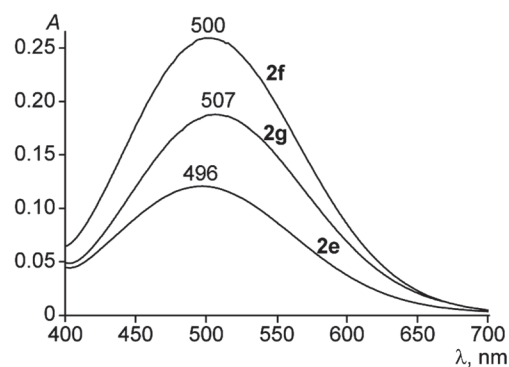


Figure 4. Normalized absorption spectra of mixtures of fluorencrownophanes **2b–g** with paraquat **6**·2PF₆ (1:10) in CH₃CN.

fluorenocrownphanes were recorded under the same conditions. The bis(fluoreno)crownphanes **2a-d** were not studied because of their very low solubility which did not allow to reach the necessary concentration of the components.

Upfield shifts compared to the chemical shifts in spectra of individual compounds were observed for the aromatic fragments of crownphanes and paraquat, as well as for fluorene H₉ protons in the ¹H NMR spectra of equimolar mixtures of fluorenocrownphanes **2e-g** with paraquat **6**·2PF₆ (Table 3, Figure 5). It clearly indicates the formation of host-guest complexes because only in this case there can be mutual shielding of the two aromatic fragments of the crownphane and the pyridyl rings of paraquat-dication **6**²⁺.^[33]

The largest upfield shifts of aromatic and H₉ proton signals are observed for crownphane **2f**. Because the magnitude of the upfield shift reflects the degree of the overlap of the opposite aromatic fragments of crownphanes and paraquat, in the first approximation it can be considered proportional to the stability of the corresponding complexes. Hence crownphane **2f** forms a more stable complex with paraquat than the **2e** and **2g**. It is important to note that the magnitudes of the shifts of H_α and H_β paraquat protons observed in case of complexes with naphthalene-containing fluorenocrownphanes **2e-g** are the largest among all crownphanes studied by us.^[17,19,21–24] An interesting fact is that the shift

of H₉ protons is almost zero for [**2e@6**]²⁺ complex. It can be caused by two reasons: the structure of this complex has significant differences from the complexes of larger crownphanes, or the shielding effect of paraquat-dication is comparable to one of a naphthalene fragment shielding H₉ protons in a non-complexed crownphane.

Conclusions

Summing up the obtained spectral data it can be concluded that fluorenocrownphanes with tri-, tetra- and pentaethylene glycols residues form relatively stable complexes with paraquat **6**·2PF₆ that are stabilized by π-π donor-acceptor interactions. The most stable complexes are formed by crownphanes with tetraethylene glycol linkers. The smallest crownphane with diethylene glycol residues does not form host-guest complexes under the conditions of the conducted experiments.

Further accumulation of experimental material, extension of the series of electrons-deficient guests and determination of the corresponding complex stability constants will allow to study the discovered dependencies and make more unambiguous conclusions about the structure of the formed complexes.

Table 3. Induced chemical shifts ($\Delta\delta$) of the aromatic protons of the crownphanes **2e-g** and paraquat **6**·2PF₆ in ¹H NMR spectra of their equimolar mixtures in CD₃CN:CDCl₃, 4:3 at 298 K.

Compound	- $\Delta\delta$								
	H _a	H _b	H _c	H ₉	H _{2,6}	H _{3,7}	H _{4,8}	H _α	H _β
2e	0.18	0.13	0.26	0.02	0.09	0.01	0.34	0.27	0.83
2f	0.22	0.22	0.36	0.23	0.01	-0.03	0.30	0.45	0.89
2g	0.12	0.11	0.23	~0.16 ¹	0.01	0.05	0.16	0.31	0.59

¹The exact determination of $\Delta\delta$ is complicated by the overlapping of the signals of H₉ protons and CH₂O groups.

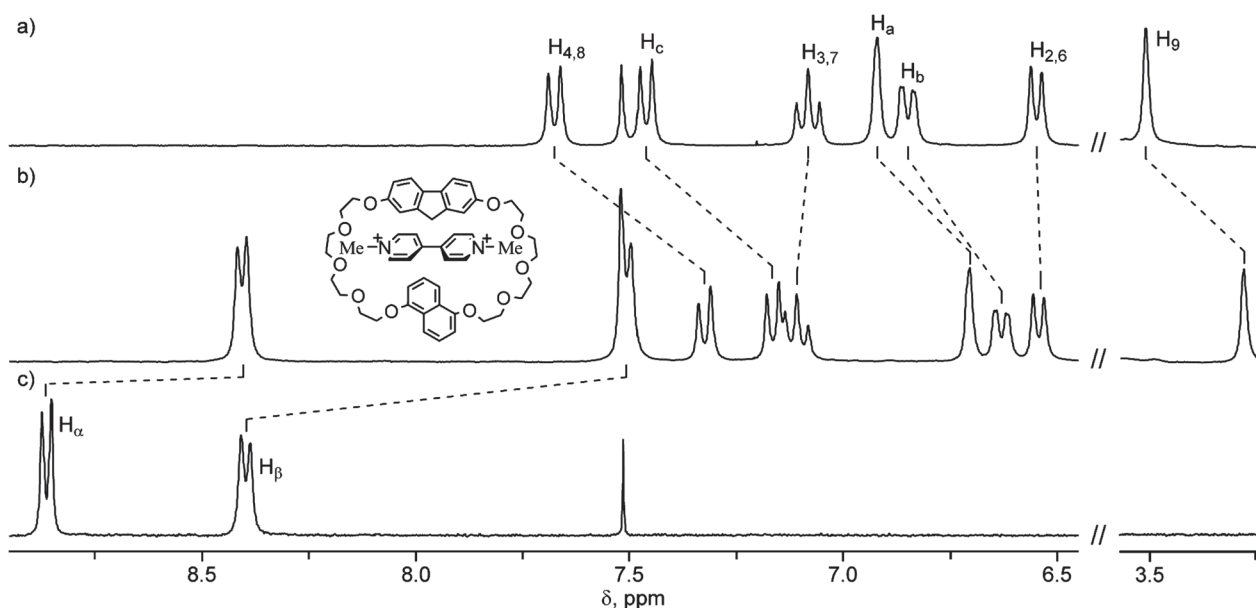


Figure 5. Parts of the ¹H NMR spectra of crownphanes **2f** (a), paraquat **6**·2PF₆ (c) and their equimolar mixture (b) in CD₃CN-CDCl₃, 4:3.

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