

ESI–MS Study of Ionization Pathways and Cation–Receptor Properties of the Iron(II) Mono– and Bis–Clathrochelates

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Dedicated to the Corresponding member of the Russian Academy of Sciences Prof. Oscar Koifman on the occasion of his 70th Birthday

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Ionization pathways of iron(II) di- and hexachloromonoclathrochelates, bis-clathrochelates and their oximehydrazonate, dimethyl- and diamine-containing macrobicyclic analogs, and the cation-receptor properties of such cage compounds towards alkali metal cations (as individual ions or their mixtures) as well as the competitive complexation in the presence of 18-crown-6 were studied by ESI-MS method. Ionization of iron(II) mono- and bis-clathrochelates under the conditions of ESI-MS experiments was found to proceed via different pathways. The macrobicyclic iron(II) oximehydrazonate as an ionic associate of the clathrochelate cation with BF_4^- anion undergoes the ionization by its heterolytic dissociation. The main pathways of ionization of the dioximate cage and bis-cage intracomplexes are substantially affected by ribbed substituents in their chelate fragments. The methyl- and amine-containing macrobicyclic complexes easily form anion-radical species by one-electron oxidation of their polyazomethine cage frameworks, whereas in the case of iron(II) dihalogenoclathrochelates the most intensive peaks correspond to their ionic associates with monocharged alkali metal cations. ESI-MS positive-mode spectrum of the iron(II) hexachloroclathrochelate contains no detectable peaks of the cationic species, which resulted either from its oxidation to the corresponding cation-radical or from its ionic associates with alkali metal cations. In the negative mode, an intensive peak assigned to the anion-radical product of one-electron reduction was observed. As ESI mass spectrometry allows maintaining precise concentrations of the complex under study and that of “cationization agents” (i.e., the compounds that promote formation of cationic species of the analyte and have been used in the case of low-ionizable complexes), the alkali metal tetraphenylborates were applied as such agents to study cation-receptor properties of the iron(II) mono- and bis-clathrochelates with ribbed substituents of the different nature towards these metal ions. Iron(II) monoclathrochelates form two types of ionic associates with such cations with both the 1:1 and 2:1 stoichiometries, which are more and less intensive in their ESI-MSs, respectively, whereas ESI-MSs of bis-cage complexes contain the intensive peaks of only 1:1 ionic associates with high affinity towards Cs^+ cation.

Keywords: Macrocyclic compounds, clathrochelates, iron complexes, ionic mass spectrometry, receptors.

Исследование путей ионизации и катион–рецепторных свойств моно– и бис–клатрохелатов железа(II) методом ESI масс–спектрометрии

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Методом ESI масс-спектрометрии были изучены пути ионизации ди- и гексахлоромонохелатов железа(II), бис-клатрохелатов железа(II) и их оксимгидразонатных макробициклических аналогов, а также катион-рецепторные свойства этих комплексов по отношению к катионам щелочных металлов.

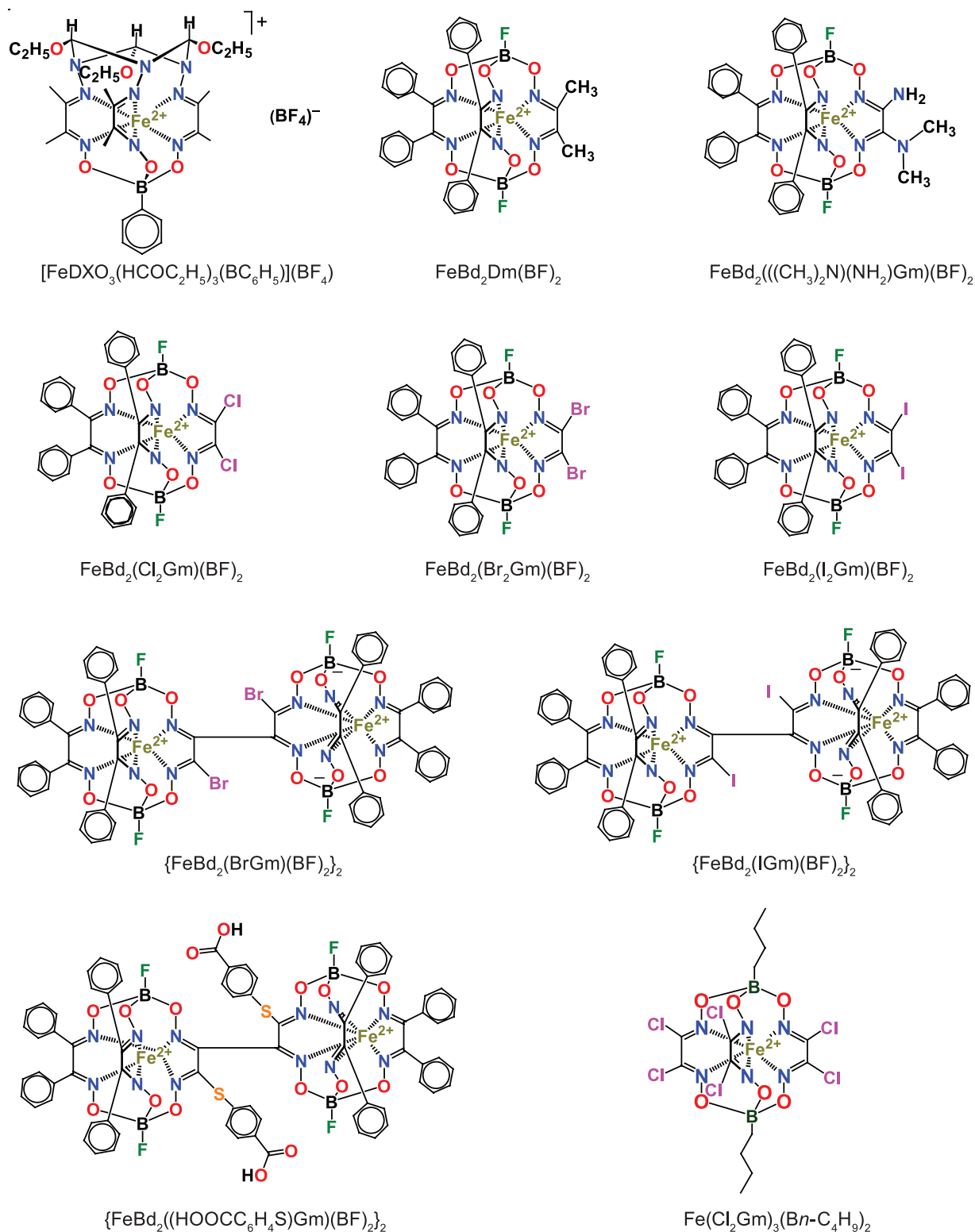
Ключевые слова: макроциклические соединения, клатрохелаты, комплексы железа, ионная масс-спектрометрия, рецепторы.

Introduction

Macrobicyclic *d*-metal tris-dioximates and tris-oximehydrazonates belong to a unique type of cage complexes (the so-called clathrochelates), with a caged transition metal ion fully encapsulated in a three-dimensional cavity of a rigid polyene macrobicyclic ligand isolating it from external factors.^[1] Biological activity of such compounds as enzymatic regulators in the systems of biosynthesis of nucleic acids has recently started to attract much attention.^[2,3] The interest of biochemists and molecular biologists in *d*-metal clathrochelates has been triggered by their high synthetic availability, (photo)chemical stability, and the activity of a series of iron(II) clathrochelates as efficient transcription inhibitors for the T7 RNA polymerase.^[3] We have recently succeeded to boost the inhibitory activity of cage compounds by direct joining of two macrobicyclic fragments with a covalent bond,^[4] while keeping all the advantages of clathrochelates as enzyme inhibitors in submicromolar range. Such iron(II) bis-clathrochelates have an incredible chemical stability and low toxicity, are easy to synthesize from available and inexpensive initial compounds, and have favorable geometry allowing introduction of up to fourteen substituents. These cage complexes have been also used as versatile guests to form strong supramolecular assemblies with blood transport proteins - serum albumins, but discriminating other globular proteins,^[5] and have been shown to affect protein uncontrolled aggregation by changing the kinetics of insulin fibrillization and reducing the amount of amyloid fibrils formed and suppressing the formation of superfibrillar species as well.^[6] This demonstrates huge potential of cage metal complexes as prospective antiviral and anticancer drug candidates (the

so-called “topological drugs”^[2-6]). The cage complexes have been also proposed^[7-28] as molecular scaffolds for the design of multicentred molecular and supramolecular systems for the target delivery of an encapsulated radionuclide as well as (radio)diagnostic and (radio)therapeutic compounds (in particular, for ¹¹B-NCT treatment^[24-28]).

The α -dioximate fragments of some mono- and trisubstituted-functionalized metal clathrochelates contain in such chelate moieties the donor *vic*-substituents (halogen atoms, amine, hydroxyl and mercapto groups) that are able to coordinate to a metal ion.^[29-31] We thus aimed on studying cation-receptor properties of halogen-containing iron(II) mono- and bis-clathrochelates (Scheme 1) having from two (FeBd₂(Cl₂Gm)(BF)₂, FeBd₂(Br₂Gm)(BF)₂, FeBd₂(I₂Gm)(BF)₂, {FeBd₂(BrGm)(BF)₂}₂ and {FeBd₂(IGm)(BF)₂}₂) to six (Fe(Cl₂Gm)₃(Bn-C₄H₉)₂) chlorine atoms in their chelate α -dioximate fragments. Note that these reactive compounds have been earlier recognized as convenient macrobicyclic precursors for their further functionalization with *N,O,C,S*-nucleophiles^[3,24,27,29-38] (including those with ionophoric, pharmacophoric and fluorophoric groups), and for the design of hybrid and multicentered compounds.^[24,27,28] Here we also describe the cation-receptor properties of their dimethyl- and diamine-containing clathrochelate analogs FeBd₂Dm(BF)₂ and FeBd₂(((CH₃)₂N)(NH₂)Gm)(BF)₂ towards alkali metal cations (as individual species or their mixtures), together with the peculiarities of ESI-MS ionization of such cage compounds; the clathrochelate oximehydrazonate [FeDXO₃(HCOC₂H₅)₃(BC₆H₅)₃](BF₄), containing a macrobicyclic cation with *m/z* 654.27 Da, was used as an internal standard. We have also studied a competitive complexation of these clathrochelates with alkali metal cations in the presence of 18-crown-6 forming stable complexes with them.



Scheme 1.

Experimental

Sodium, potassium, cesium and rubidium tetraphenylborates as well as organic solvents (analytical grade) were obtained commercially (SAF). The iron(II) mono- and bis-clathrochelates under study were obtained as described previously.^[4,32,37-42]

ESI-MS were obtained on a Bruker MicrOTOF-Q spectrometer equipped with an Apollo II electrospray ionization source with an ion funnel. The instrumental parameters were

as follows: scan range m/z 200–2200 Da, dry gas: nitrogen, temperature 200 °C, transfer time 120 ps. The sample was infused at a flow rate of 3 $\mu\text{L}\cdot\text{min}^{-1}$. Before each run, the spectrometer was calibrated externally with the Tunemix™ mixture (Bruker Daltonik, Germany) in a quadratic regression mode. The mass accuracy for the calibration was better than 5 ppm, together with the true isotopic pattern (using a SigmaFit) enabling an unambiguous conformation of the elemental composition of the clathrochelate complexes studied.

Table 1. ESI-MS data for the iron(II) dihalogenoclathrochelates in the presence of equimolar amounts of alkali metal cations.

	System M+Na ⁺			System M+K ⁺			System M+Rb ⁺			System M+Cs ⁺		
	[M] ⁺	[M+Na ⁺] ⁺	[2M+Na ⁺] ⁺	[M] ⁺	[M+K ⁺] ⁺	[2M+K ⁺] ⁺	[M] ⁺	[M+Rb ⁺] ⁺	[2M+Rb ⁺] ⁺	[M] ⁺	[M+Cs ⁺] ⁺	[2M+Cs ⁺] ⁺
FeBd₂(Cl₂Gm)(BF)₂												
Intensity (a.u.)	125	42065	5590	929	9220	1727	4639	5777	663	2033	35115	1637
<i>I</i> rel. (%)	0.3	100	11.7	10.08	100	18.73	80.31	100	11.48	5.79	100	4.66
<i>I</i> / <i>I</i> sum. (%)	0.26	88.04	11.70	7.82	77.64	14.54	41.87	52.14	5.98	5.24	90.54	4.22
<i>m/z</i> (Da)	747.04	769.02	785	747.04	785.00	1533.08	747.04	830.95	1577.99	747.04	878.93	1625.96
FeBd₂(Br₂Gm)(BF)₂												
Intensity (a.u.)	223	10741	1781	579	7993	1044	1482	6817	895	564	16652	1595
<i>I</i> rel. (%)	2.07	100	16.58	7.24	100	13.06	21.74	100	13.13	3.39	100	9.58
<i>I</i> / <i>I</i> sum. (%)	1.75	84.28	13.97	6.02	83.12	10.86	16.12	74.15	9.73	3.00	88.52	8.48
<i>m/z</i> (Da)	835.95	858.92	1693.87	835.95	874.89	1710.85	835.95	920.85	1756.77	835.95	968.85	1804.80
FeBd₂(I₂Gm)(BF)₂												
Intensity (a.u.)	144	3339	323	181	2080	256	80	3215	352	159	13277	1242
<i>I</i> rel. (%)	4.31	100	9.68	8.71	100	12.3	2.49	100	10.94	1.2	100	9.36
<i>I</i> / <i>I</i> sum. (%)	3.78	87.73	8.49	7.19	82.64	10.17	2.19	88.15	9.65	1.08	90.46	8.46
<i>m/z</i> (Da)	931.29	952.89	1882.82	931.29	968.87	1898.81	931.29	1014.82	1944.72	931.29	1062.82	1992.76

Table 2. ESI-MS data for the iron(II) bis-clathrochelates in the presence of equimolar amounts of alkali metal cations.

	[M] ⁺	[M+Na ⁺] ⁺	[M+K ⁺] ⁺	[M+Rb ⁺] ⁺	[M+Cs ⁺] ⁺	
{FeBd₂(IGm)(BF)₂}₂						
Intensity (a.u.)		5421826	44465056	14798896	4853671	194648607
^a <i>I</i> rel. (%)		3	23	8	2	100
^b <i>I</i> / <i>I</i> sum. (%)		2	17	6	2	74
<i>m/z</i> (Da)		1354	1377	1393	1439	1487
{FeBd₂(BrGm)(BF)₂}₂						
Intensity (a.u.)		9650692	21359035	34330723	69080510	159036327
^a <i>I</i> rel. (%)		6	13	22	43	100
^b <i>I</i> / <i>I</i> sum. (%)		3	7	12	24	54
<i>m/z</i> (Da)		1512.1	1535.1	1551.1	1597.0	1645.0
{FeBd₂((HOCC₆H₄S)Gm)(BF)₂}₂						
Intensity (a.u.)		3048045	1158679	10354901	2711655	25094479
^a <i>I</i> rel. (%)		12	5	41	11	100
^b <i>I</i> / <i>I</i> sum. (%)		7	3	24	6	59
<i>m/z</i> (Da)		1658.3	1681.3	1697.2	1744.3	1791.2

^aIntensity relative to most intensive peak;^bRelative intensity to the sum of all peaks observed.

1 mM solutions of the complexes studied and alkali metal tetraphenylborates were obtained by a dissolution of their weighted amounts in pure acetonitrile for 20 min with an ultrasonic dispersion followed by a centrifugation.

Four series of the ESI-MS experiments were performed (a-d, below). (a) The mass spectra of the acetonitrile solutions of these complexes were obtained at their concentrations $5 \cdot 10^{-5}$ mol·l⁻¹. (b) The spectral data were collected for the mixtures of 1mM acetonitrile solution of the cage complex under study (25 μl), 1mM acetonitrile solution of the corresponding alkali metal tetraphenylborate (25 μl) and acetonitrile (450 μl). The obtained solutions with equimolar concentrations of the components equal to $5 \cdot 10^{-5}$ mol·l⁻¹ underwent an ultrasonic dispersion and then were studied at the same experimental conditions; the

corresponding ESI-MS data are summarized in Tables 1 and 2. (c) The ESI-MSs from the mixtures of 1mM acetonitrile solution of the clathrochelate under study (25 μl) and 1mM acetonitrile solutions of the sodium, potassium, rubidium and cesium tetraphenylborates (25 μl each) and acetonitrile (375 μl) were recorded. The solutions obtained with equimolar concentrations of the components equal to $5 \cdot 10^{-5}$ mol·l⁻¹ were treated and studied as described above; the corresponding ESI-MS data are summarized in Table 3. (d) Finally, the concurrent complexation of the mono- and bis-clathrochelates FeBd₂(Br₂Gm)(BF)₂, {FeBd₂(Gm-S-C₆H₄-COOH)(BF)₂}₂ and {FeBd₂(BrGm)(BF)₂}₂ with alkali metal cations in the presence of 18-crown-6 was studied. Two microsyringes (one containing 50 μM acetonitrile solution of 18-crown-6, the other a mixture of acetonitrile solutions of the clathrochelate under study,

Table 3. ESI-MS data for the iron(II) clathrochelates in the presence of equimolar mixture of alkali metal cations.

Compound	[M] ⁺	[M+Na ⁺] ⁺	[M+K ⁺] ⁺	[M+Rb ⁺] ⁺	[M+Cs ⁺] ⁺	[2M+Na ⁺] ⁺	[2M+K ⁺] ⁺	[2M+Rb ⁺] ⁺	[2M+Cs ⁺] ⁺
FeBd ₂ (Cl ₂ Gm)(BF) ₂									
Intensity (a.u.)	1787	36652	12236	14088	44738	6508	1414	1302	1944
<i>I</i> rel. (%)	4	82	27	31	100	15	3	3	4
<i>I</i> / <i>I</i> sum. (%)	1.5	30.4	10.1	11.7	37.1	5.4	1.2	1.1	1.6
<i>m/z</i> (Da)	746	769	785	832.9	878	1517.1	1533	1579	1627
FeBd ₂ (Br ₂ Gm)(BF) ₂									
Intensity (a.u.)	669	17058	7777	10721	29096	1872	954	909	1667
<i>I</i> rel. (%)	2.3	58.64	26.73	36.85	100	6.43	3.28	3.12	5.74
<i>I</i> / <i>I</i> sum. (%)	0.9	24.1	11.0	15.2	41.1	2.6	1.3	1.3	2.4
<i>m/z</i> (Da)	836	859	875	921	969	1695	1711	1757	11805
FeBd ₂ (I ₂ Gm)(BF) ₂									
Intensity (a.u.)	5088	11616	17176	16601	33890	1207	2495	2721	4476
<i>I</i> rel. (%)	15	34	51	49	100	4	7	8	13
<i>I</i> / <i>I</i> sum. (%)	5.3	12.2	18.0	17.4	35.6	1.3	2.6	2.9	4.7
<i>m/z</i> (Da)	930	953	969	1015	1063	1883	1899	1945	1993
FeBd ₂ ((CH ₃) ₂ N) (NH ₂ Gm)(BF) ₂									
Intensity (a.u.)	23488991	26358082	10313148	5359212	65273469	17746021	7985475	2940943	14866928
<i>I</i> rel. (%)	35.99	40.38	15.80	8.21	100	27.19	12.23	4.51	22.78
<i>I</i> / <i>I</i> sum. (%)	13.47	15.12	5.92	3.07	37.44	10.18	4.58	1.69	8.53
<i>m/z</i> (Da)	736	759	775	821	869	1495	1511	1557	1605

Table 4. ESI-MS data for the dibromine-containing iron(II) mono- and bis-clathrochelates and their equimolar mixture in the presence of equimolar amounts of the pair of cations Na⁺/Cs⁺ and the clathrochelate oximehydrazone cation [FeDXO₃(HCO₂H₃)(BC₆H₅)](BF₄) as an internal standart.

	[FeDXO ₃ (HCO ₂ H ₃)(BC ₆ H ₅)](BF ₄)	[M + Na ⁺] ⁺	[M + Cs ⁺] ⁺	[M+Cs ⁺⁺ CH ₃ OH] ⁺	[2M+Na ⁺] ⁺	[2M+Cs ⁺] ⁺
FeBd ₂ (Br ₂ Gm)(BF) ₂						
Intensity (a.u.)	310950295	40319282	426289171	329399064	6599297	42410201
<i>I</i> rel. (%)	72.94	9.46	100.00	77.27	1.55	9.95
<i>m/z</i> (Da)	654.2763	858.9437	968.8625	1001.0472	1694.9327	1804.8293
{FeBd ₂ (BrGm)(BF) ₂ } ₂						
Intensity (a.u.)	134745266	20408569	224523368	75100143		
<i>I</i> rel. (%)	60.01	9.09	100.00	33.45		
<i>m/z</i> (Da)	654.2744	1535.0809	1645.0029	1677.1763		
FeBd ₂ Br ₂ Gm(BF) ₂ + {FeBd ₂ BrGm(BF) ₂ } ₂						
Intensity (a.u.)	85172319	14138276 12580616	71549421 96842853	63493095 28829004	1228349	8982712
<i>I</i> rel. (%)	87.95	14.60 12.99	73.88 100.00	65.56 29.77	1.27	9.28
<i>m/z</i> (Da)	654.2764	858.9437 1535.0803	968.8624 1645.0028	1001.0472 1677.1765	1694.9332	8982712

cesium tetraphenylborate, and oximehydrazone clathrochelate [FeDXO₃(HCO₂H₃)(BC₆H₅)](BF₄) as an internal standard, at the same concentrations) were connected with input unit of the sample into a spectrometer by a mixer. The rates of the addition of these solutions to the mixer were controlled in such a manner that their volumes in the mixer were constant; the spectra were recorded at each step and the data obtained are summarized in Table 5.

Results and Discussion

In contrast to MALDI-TOF mass spectrometry, with the concentration of an analyte depending on the coordinates of the laser-irradiated point on a target, the ESI mass spectrometry allows maintaining precise concentrations of the complex under study and those of the "cationization

Table 5. ESI-MS data for the mixtures of the clathrochelate $\text{FeBd}_2\text{Br}_2\text{Gm}(\text{BF}_2)_2 - \text{Cs}^+ - 18\text{-crown-6}$.

Clathrochelate-to-18-crown-6 molar ratio	without 18-crown-6	1	2	6	8
$[\text{M} + \text{Cs}^+]^+$					
<i>I</i> rel. (%)	20.12	23.01	6.01	0.14	0.08
<i>m/z</i> (Da)	968.8634	968.8636	968.8635	968.8595	968.8636
$[\text{M} + \text{Cs}^+ + \text{CH}_3\text{OH}]^+$					
<i>I</i> rel. (%)	32.97	31.74	13.86	5.82	2.15
<i>m/z</i> (Da)	1001.0485	1001.0485	1001.0484	1001.0484	1001.0485
$[2\text{M} + \text{Cs}^+]^+$					
<i>I</i> rel. (%)	24.09	21.31	3.76	0.99	0.85
<i>m/z</i> (Da)	1804.8643	1804.8644	1804.8645	1804.8648	1804.8647
$[18\text{-crown-6} + \text{Cs}^+]^+$					
<i>I</i> rel. (%)	0.00	7.80	48.77	84.87	100.00
<i>m/z</i> (Da)	397.0653	397.0606	397.0653	397.0653	397.0653

agents" (*i.e.*, the compounds that promote formation of cationic species of an analyte and are used in the case of low-ionizable complexes).

We have used the alkali metal tetraphenylborates as the cationization agents to study cation–receptor properties of the iron(II) mono- and bis-clathrochelates with ribbed substituents of the different nature towards various alkali metal cations.

In the absence of alkali metal tetraphenylborates, the main pathway of the ionization of the alkyl- and amine-containing clathrochelates $\text{FeBd}_2\text{Dm}(\text{BF}_2)_2$ and $\text{FeBd}_2(((\text{CH}_3)_2\text{N})(\text{NH}_2)\text{Gm})(\text{BF}_2)_2$ under the ESI-MS experimental conditions is their one-electron oxidation resulting in the molecular ions $[\text{M}]^{+\bullet}$. When the equimolar or excess amounts of these cationization agents are present, the corresponding spectra show intensive peaks of the ionic associates between the cage complexes and alkali metal cations (Table 1).

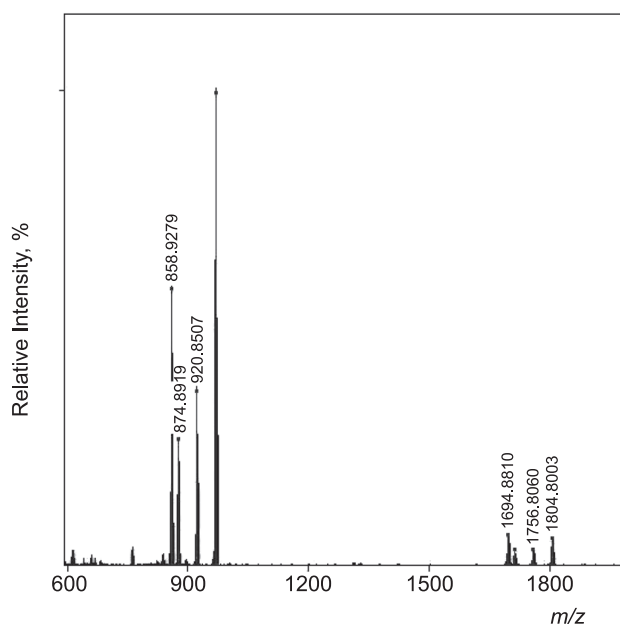


Figure 1. Positive range of ESI-MS for the clathrochelate $\text{FeBd}_2(\text{Br}_2\text{Gm})(\text{BF}_2)_2$ in the presence of equimolar amounts of sodium, potassium, rubidium and cesium tetraphenylborates.

The initial ESI-MSs of the iron(II) dihalogenoclathrochelates contain the low-intensive peaks of their ionic associates with impurity Na^+ and K^+ cations. In the presence of the alkali metal tetraphenylborates, the intensities of these peaks increase; ESI-MS of the acetonitrile solution of dibromoclathrochelate $\text{FeBd}_2(\text{Br}_2\text{Gm})(\text{BF}_2)_2$ with the equimolar amounts of sodium, potassium, rubidium and cesium tetraphenylborates is shown in Figure 1. This spectrum contains two groups of the most intensive peaks: (i) with *m/z* from 858 to 969 Da, assigned to the monocharged ionic associates of this cage complex with alkali metal cations $[\text{M} + \text{Cat}]^+$, and (ii) with *m/z* from 1695 to 1804 Da, assigned to the ionic associates between two cage species and one alkali metal cation $[2\text{M} + \text{Cat}]^+$. In the spectra of other iron(II) clathrochelates and bis-clathrochelates studied, the peaks of their ionic associates with alkali metal cations also dominate, whereas the signals of the protonated cage complexes $[\text{M} + \text{H}^+]^+$ are either not observed or low-intensive.

The ESI-MS spectrum of the hexachloroclathrochelate $\text{Fe}(\text{Cl}_2\text{Gm})_3(\text{Bn}-\text{C}_4\text{H}_9)_2$, recorded in positive mode, contains no peaks of the cationic species, resulted either from compound oxidation or its association with alkali metal cations. In the negative mode, the intensive peak $[\text{M}]^{\bullet-}$ of the anion-radical macrobicyclic product of clathrochelate one-electron reduction was observed.

The spectral data of the cage complexes $\text{FeBd}_2(((\text{CH}_3)_2\text{N})(\text{NH}_2)\text{Gm})(\text{BF}_2)_2$, $\text{FeBd}_2(\text{Cl}_2\text{Gm})(\text{BF}_2)_2$, $\text{FeBd}_2(\text{Br}_2\text{Gm})(\text{BF}_2)_2$ and $\text{FeBd}_2(\text{I}_2\text{Gm})(\text{BF}_2)_2$ clearly showed the selectivity of their binding with alkali metal cations: the intensities of the corresponding peaks decrease in the series $\text{Cs}^+ > \text{Na}^+ > \text{Rb}^+ > \text{K}^+$ (Figure 2). It should be noted that for all the iron(II) dihalogenoclathrochelates studied, the most intensive peaks correspond to their ionic associates with Cs^+ and Na^+ ; no linear relation of their intensities vs the ionic radius of an alkali metal cation is observed. The parent ESI-MS for the iron(II) bis-clathrochelates contain no peaks of their protonated $[\text{M} + \text{H}^+]^+$ or cation-radical oxidized $[\text{M}]^{+\bullet}$ forms; these, however, may be too weak to be observed.

Note that in the case of a mixture of equimolar amounts of alkali metal ions, for all these complexes the peaks of their ionic associates with Cs^+ ion are the most intensive (Figures 2 and 3). Another peculiarity of the spectra of the

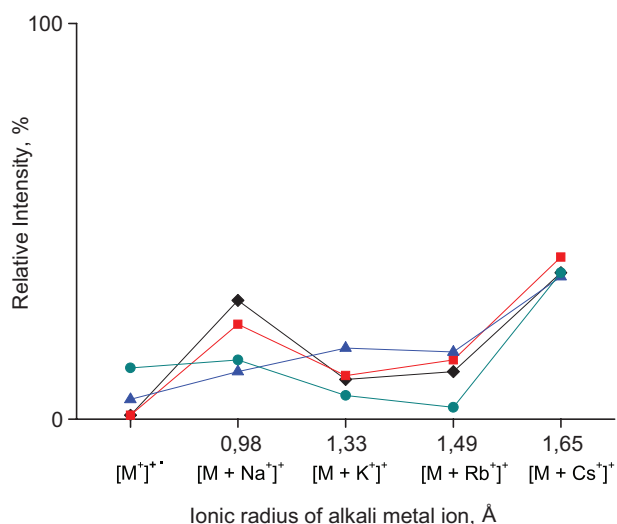


Figure 2. Plots of the relative intensities of the peaks assigned to the $[M + \text{Cat}]^+$ cations vs the type of an alkali metal cation for macrobicyclic compounds $\text{FeBd}_2(\text{Cl}_2\text{Gm})(\text{BF})_2$ (\blacklozenge), $\text{FeBd}_2(\text{Br}_2\text{Gm})(\text{BF})_2$ (\blacksquare), $\text{FeBd}_2(\text{I}_2\text{Gm})(\text{BF})_2$ (\blacktriangle) and $\text{FeBd}_2(\text{((CH}_3)_2\text{N)(NH}_2\text{)Gm)(BF)}_2$ (\bullet); the data for the molecular ion $[M]^+$ are also included.

iron(II) complexes studied is the presence of $[M + \text{Cat}]^+$ and $[2M + \text{Cat}]^+$ peaks of the ionic associates with clathrochelates-to-alkali metal cation stoichiometric ratios of 1:1 and 2:1, whereas those of their bis-clathrochelate analogs contain intensive peaks of 1:1 associates only.

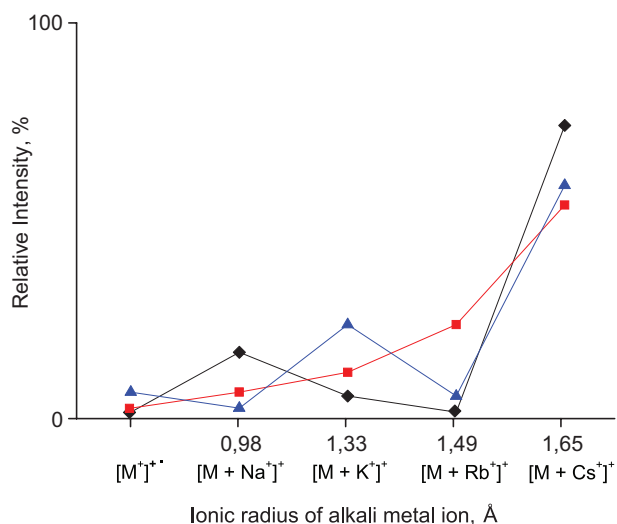


Figure 3. Plots of the relative intensities of the peaks assigned to the $[M + \text{Cat}]^+$ cations vs the type of an alkali metal cation for the bis-clathrochelates $\{\text{FeBd}_2(\text{I}_2\text{Gm})(\text{BF})_2\}_2$ (\blacklozenge), $\{\text{FeBd}_2(\text{BrGm})(\text{BF})_2\}_2$ (\blacksquare) and $\{\text{FeBd}_2(\text{(HOOC}_6\text{H}_4\text{S)Gm)(BF)}_2\}_2$ (\blacktriangle); the data for the molecular ion $[M]^+$ are also included.

The observed dependence of a selectivity of the binding of these alkali metal cations in their equimolar mixture by the iron(II) mono- and bis-clathrochelates on the nature of their ribbed substituents is the result of fine effects of cation recognition by these cage complexes.

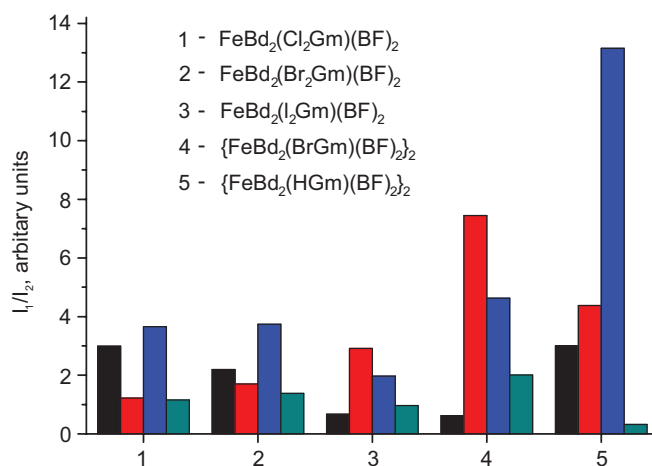


Figure 4. Relative intensities of the peaks assigned to the ionic associates $[M + \text{Cat}]^+$ for the iron(II) mono- and bis-clathrochelates and the pairs of alkali metal cations Na^+/K^+ (\blacksquare); Cs^+/Na^+ (\blacksquare); Cs^+/K^+ (\blacktriangle) and Rb^+/K^+ (\bullet).

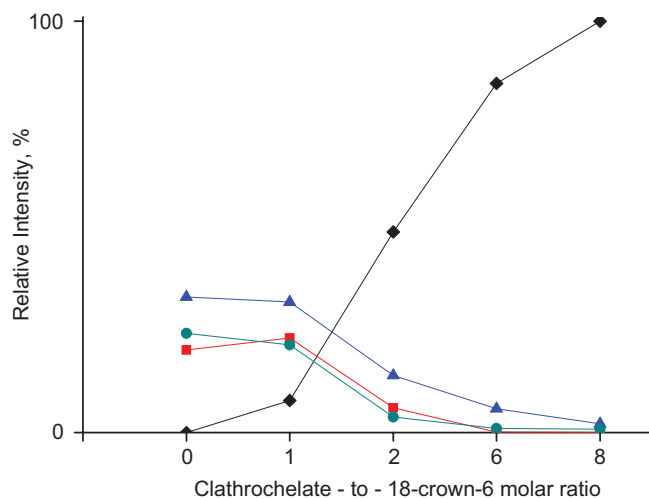


Figure 5. Plots of the intensities of the peaks of different ionic associates $\{[M + \text{Cs}]^+$ (\blacksquare), $[M + \text{Cs} + \text{CH}_3\text{OH}]^+$ (\blacktriangle) and $[2M + \text{Cs}]^+$ (\bullet) of Cs^+ cation with the clathrochelate $\text{FeBd}_2(\text{Br}_2\text{Gm})(\text{BF})_2$ and with 18-crown-6 (\blacklozenge) as a concurrent ligand vs the clathrochelate-to-18-crown-6 molar ratio.

A stepwise addition of 18-crown-6 as a competitive ligand to the acetonitrile solution of the clathrochelate $\text{FeBd}_2(\text{Br}_2\text{Gm})(\text{BF})_2$ with the equimolar amount of cesium tetraphenylborate caused the substantial changes in the intensities of the peaks in its ESI-MS: the intensity of the peak with m/z 397 Da assigned to the cationic species $[\text{18-crown-6} + \text{Cs}]^+$ increases, whereas for those assigned to the ionic associates of this clathrochelate with Cs^+ cation correspondingly decreases. New peaks of either the molecular clathrochelate ions or their fragments do not appear in the spectrum. Therefore, 18-crown-6 as a competitive ligand plays a role of the so-called “anticationization agent” by suppressing the main pathway of the ionization of the cage complexes, *i.e.* the formation of their ionic associates with alkali metal cations.

Conclusions

We have studied the peculiarities of the ionization of different iron(II) mono- and bis-clathrochelates under the conditions of ESI-MS experiments. The macrobicyclic iron(II) oximehydrazonate as an ionic associate of the macrobicyclic cation with BF_4^- anion undergoes the ionization by its heterolytic dissociation. The main pathways of ionization of the dioximate cage and bis-cage intracomplexes are substantially affected by ribbed substituents in their chelate fragments. The alkyl- and amine-containing macrobicyclic complexes easily form anion-radical species $[\text{M}]^{\cdot-}$ by one-electron oxidation of their polyazomethine cage frameworks, whereas in the case of iron(II) dihalogenoclathrochelates the most intensive peaks belong to their ionic associates with monocharged alkali metal cations. Positive-mode ESI-MS spectrum of the iron(II) hexachloroclathrochelate have not shown detectable peaks of the cationic species, resulted either from its oxidation to the corresponding cation-radical or from the formation of ionic associates with alkali metal cations. An intensive peak observed in the negative mode was assigned to the anion-radical product $[\text{M}]^{\cdot-}$ of clathrochelate one-electron reduction. The iron(II) monoclathrochelates form two types of ionic associates with alkali metal cations with both the 1:1 and 2:1 stoichiometries. The ESI-MS of the bis-cage complexes contains intensive peaks of only 1:1 ionic associates with high affinity towards Cs^+ cation.

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References

- Voloshin Y.Z., Kostromina N.A., Krämer R. *Clathrochelates: Synthesis, Structure and Properties*, Elsevier, Amsterdam, **2002**.
- Voloshin Y., Varzatskii O., Shul'ga S., Novikov V., Belov A., Makarenko I., Dubey I., Krivorotenko D., Negrutska V., Zhizhin K., Kuznetsov N., Bubnov Y. *Proc. 10th EURO-BIC*, Thessaloniki, Greece, 22-26 June, **2010**, p. 29-38.
- Novikov V.V., Varzatskii O.A., Negrutska V.V., Bubnov Y.N., Palchykovska L.G., Dubey I.Y., Voloshin Y.Z. *J. Inorg. Biochem.* **2013**, *124*, 42-45.
- Varzatskii O.A., Novikov V.V., Shulga S.V., Belov A.S., Vologzhanina A.V., Negrutska V.V., Dubey I.Y., Bubnov Y.N., Voloshin Y.Z. *Chem. Commun.* **2014**, *50*, 3166-3168.
- Losytskyy M.Y., Kovalska V.B., Varzatskii O.A., Sergeev A.M., Yarmoluk S.M., Voloshin Y.Z. *J. Fluoresc.* **2013**, *23*, 889-895.
- Kovalska V.B., Losytskyy M.Yu., Varzatskii O.A., Cherepanov V.V., Voloshin Y.Z., Mokhir A.A., Yarmoluk S.M., Volkov S.V. *J. Bioorg. Med. Chem.* **2014**, 1883-1888.
- Smith S.V., Harrowfield J.M., Di Bartolo N.M., Sargeson A.M., PCT Int. Appl. WO 00 40, 585 (Cl. C07D487/08) Publ. 13.07.2000.
- Donnelly P.S., Harrowfield J.M., Skelton B.W., White A.H. *Inorg. Chem.* **2000**, *39*, 5817-5830.
- Donnelly P.S., Harrowfield J.M. *J. Chem. Soc., Dalton Trans.* **2002**, 906-913.
- Smith S.V. *J. Inorg. Biochem.* **2004**, *98*, 1874-1901.
- Di Bartolo N., Sargeson A.M., Smith S.V. *Org. Biomol. Chem.* **2006**, *4*, 3350-3357.
- Voss S.D., Smith S.V., Di Bartolo N., McIntosh L.J., Cyr E.M., Bonab A.A., Carter E.A., Fischman A.J., Treves S.T., Gillies S.D., Sargeson A.M., Huston J.S., Packard A.B. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 17489-17493.
- Cai H., Fissekis J., Conti P.S. *Dalton Trans.* **2009**, 5395-5400.
- Ma M.T., Karas J.A., White J.M., Scanlon D., Donnelly P.S. *Chem. Commun.* **2009**, 3237-3239.
- Cai H., Li Z., Huang C.-W., Park R., Shahinian A.H., Conti P.S. *Nucl. Med. Biol.* **2010**, *37*, 57-65.
- Dittrich B., Harrowfield J.M., Koutsantonis G.A., Nealon G.L. *Dalton Trans.* **2010**, *39*, 3433-3448.
- Liu S., Li Z., Yap L.-P., Huang C.-W., Park R., Conti P.S. *Chem. Eur. J.* **2011**, *17*, 10222-10225.
- Ma M.T., Neels O.C., Denoyer D., Roselt P., Karas J.A., Scanlon D.B., White J.M., Hicks R.J., Donnelly P.S. *Bioconjugate Chem.* **2011**, *22*, 2093-2103.
- Ma M.T., Cooper M.S., Paul R.L., Shaw K.P., Karas J.A., Scanlon D., White J.M., Blower P.J., Donnelly P.S. *Inorg. Chem.* **2011**, *50*, 6701-6710.
- Chen K., Wang X., Lin W.-Y., Shen C. K.-F., Yap L.-P., Hughes L.D., Conti P.S. *ACS Med. Chem. Lett.* **2012**, *3*, 1019-1023.
- Qin C.-J., James L., Chartres J.D., Alcock L.J., Davis K.J., Willis A.C., Sargeson A.M., Bernhardt P.V., Ralph S.F. *Inorg. Chem.* **2011**, *50*, 9131-9140.
- Bernhardt P.V., Font H., Gallego C., Martínez M., Rodríguez C. *Inorg. Chem.* **2012**, *51*, 12372.
- Voloshin Y.Z., Varzatskii O.A., Bubnov Y.N. *Russ. Chem. Bull.* **2007**, *56*, 577-605.
- Voloshin Y.Z., Varzatskii O.A., Zhizhin K.Y., Kuznetsov N.T., Bubnov Y.N. *Russ. Chem. Bull.* **2006**, *55*, 22-25.
- Voloshin Y.Z., Erdyakov S.Y., Makarenko I.G., Lebed E.G., Potapova T.V., Svidlov S.V., Starikova Z.A., Pol'shin E.V., Gurskii M.E., Bubnov Y.N. *Rus. Chem. Bull.* **2007**, *56*, 1787-1794.
- Erdyakov S.Y., Voloshin Y.Z., Makarenko I.G., Lebed E.G., Potapova T.V., Ignatenko A.V., Vologzhanina A.V., Gurskii M.E., Bubnov Y.N. *Inorg. Chem. Commun.* **2009**, *12*, 135-139.
- Kuznetsov N.T., Belaya I.G., Dolganov A.V., Zelinsky G.E., Matveev E.Y., Zhizhin K.Y., Voloshin Y.Z., Bubnov Y.N. *Russ. Chem. Bull.* **2011**, *60*, 2518-2521.
- Svidlov S.V., Varzatskii O.A., Potapova T.V., Vologzhanina A.V., Bukalov S.S., Leites L.A., Voloshin Y.Z., Bubnov Y.N. *Inorg. Chem. Commun.* **2014**, *43*, 142-145.
- Voloshin Y.Z., Varzatskii O.A., Kochubey D.I., Vorontsov I.I., Bubnov Y.N. *Inorg. Chim. Acta* **2009**, *362*, 149-158.
- Vershinin M.A., Burdukov A.B., Boguslavskii E.G., Pervukhina N.V., Kuratieva N.V., Eltsov I.V., Reznikov V.A., Varzatskii O.A., Voloshin Y.Z., Bubnov Y.N. *Inorg. Chim. Acta* **2011**, *366*, 91-97.
- Belov A.S., Prikhod'ko A.I., Novikov V.V., Vologzhanina A.V., Bubnov Y.N., Voloshin Y.Z. *Eur. J. Inorg. Chem.* **2012**, 4507-4514.
- Voloshin Y.Z., Zavodnik V.E., Varzatskii O.A., Belsky V.K., Palchik A.V., Strizhakova N.G., Vorontsov I.I., Antipin M.Y. *Dalton Trans.* **2002**, 1193-1202.
- Voloshin Y.Z., Varzatskii O.A., Belov A.S., Starikova Z.A., Suponitsky K.Y., Novikov V.V., Bubnov Y.N. *Inorg. Chem.* **2008**, *47*, 2155-2167.
- Varzatskii O.A., Voloshin Y.Z., Korobko S.V., Shulga S.V., Krämer R., Belov A.S., Vologzhanina A.V., Bubnov Y.N. *Polyhedron* **2009**, *28*, 3431-3438, and references cited therein.
- Voloshin Y.Z., Varzatskii O.A., Belov A.S., Starikova Z.A., Dolganov A.V., Novikov V.V., Bubnov Y.N. *Inorg. Chim. Acta* **2011**, *370*, 322-332.

36. Voloshin Y.Z., Belaya I.G., Belov A.S., Platonov V.E., Maksimov A.M., Vologzhanina A.V., Starikova Z.A., Dolganov A.V., Novikov V.V., Bubnov Y.N. *Dalton Trans.* **2012**, 41, 737-746, and references cited therein.
37. Belaya I.G., Zelinskii G.E., Belov A.S., Varzatskii O.A., Novikov V.V., Dolganov A.V., Kozłowski H., Szyrwił Ł., Bubnov Y.N., Voloshin Y.Z. *Polyhedron* **2012**, 40, 32-39.
38. Voloshin Y.Z., Varzatskii O.A., Palchik A.V., Polshin E.V., Maletin Y.A., Strizhakova N.G., *Polyhedron* **1998**, 17, 4315-4326.
39. Voloshin Y.Z., Stash A.I., Varzatskii O.A., Belsky V.K., Maletin Y.A., Strizhakov N.G., *Inorg. Chim. Acta* **1999**, 284, 180-190.
40. Voloshin Y.Z., Varzatskii O.A., Kron T.E., Belsky V.K., Zavodnik V.E., Palchik A.V. *Inorg. Chem.* **2000**, 39, 1907-1918.
41. Burdukov A.B., Vershinin M.A., Pervukhina N.V., Voloshin Y.Z., Varzatskii O.A. *Russ. Chem. Bull.* **2006**, 55, 1982-1988.
42. Voloshin Y.Z., Belov A.S., Varzatskii O.A., Shul'ga S.V., Stuzhin P.A., Starikova Z.A., Lebed E.G., Bubnov Y.N. *Dalton Trans.* **2012**, 41, 921-928.

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