

Facile Synthesis of 15<sup>2</sup>-Carboxamides of Methyl Pheophorbide *a*Nikita O. Dugin,<sup>a</sup> Maria G. Zavialova,<sup>a</sup> Roman A. Novikov,<sup>b</sup> Vladimir P. Timofeev,<sup>b</sup> Alexander Yu. Misharin,<sup>a</sup> and Gelii V. Ponomarev<sup>a@</sup><sup>a</sup>Orekhovich Institute of Biomedical Chemistry RAMS, 119121 Moscow, Russia<sup>b</sup>Engelhardt Institute of Molecular Biology RAS, 119991 Moscow, Russia

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The novel synthesis of chlorin 15<sup>2</sup>-carboxamides from methyl pheophorbide, which passed without cleavage of exocycle *E*, is developed. The combination of this approach together with known method of opening exocycle *E* by amines allows easy preparation of 13<sup>1</sup>- and 15<sup>2</sup>-carboxamides of chlorin *e*<sub>6</sub> and methyl pheophorbide *a*, comprising various substituents in amide moieties. Two examples (where substituents are either benzyl-, or hexadecyl-) are presented.

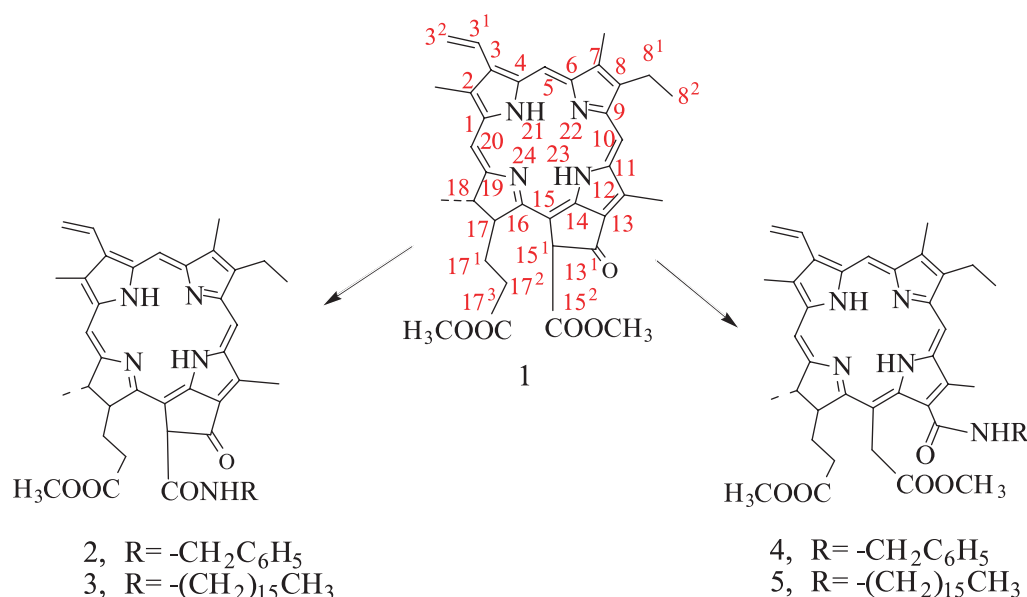
**Keywords:** Chlorin *e*<sub>6</sub>, methyl pheophorbide *a*, amidation

Tetrapyrrol macrocycles of chlorin family are widely used as optical and fluorescent sensors in various biological studies, as well as sensitizers for tumor photodynamic therapy.<sup>[1-7]</sup> Chemical modification of periphery substituents allows significantly change physical, chemical, spectral and photochemical properties of macrocycle and improve its solubility in various media, affinity to specific targets and photodynamic applications. A large number of chlorin derivatives comprising macrocycle modified with polyamines, amino acids, peptides,<sup>[8-14]</sup> carbohydrates,<sup>[15]</sup> steroids and lipids<sup>[16-18]</sup> have been synthesized and used. Therefore development of new convenient methods for introducing of substituents in various positions of chlorin macrocycle is of importance.

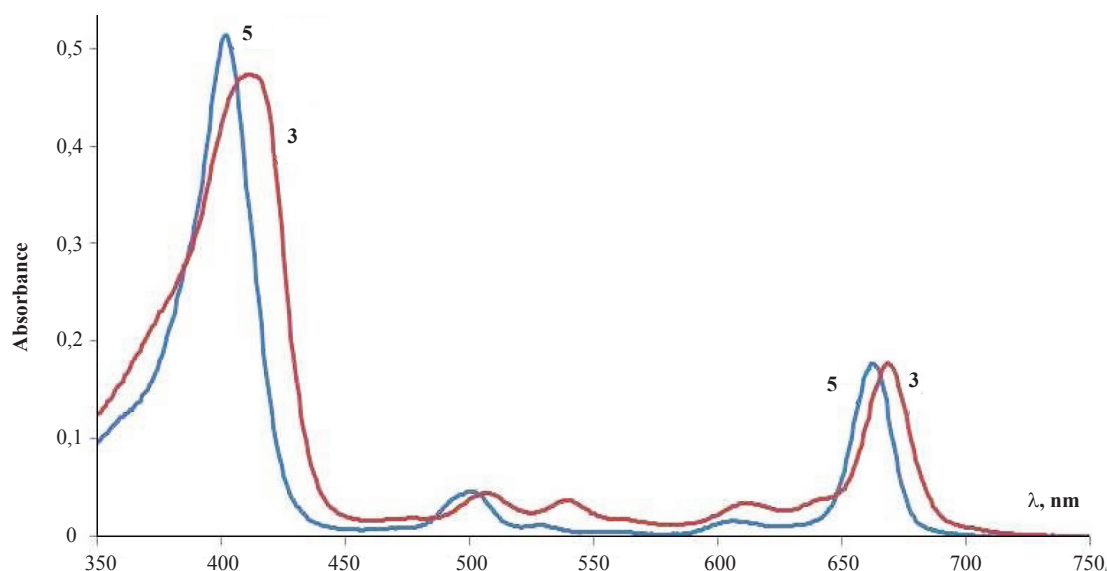
Herein we present a simple method for synthesis of chlorin 15<sup>2</sup>-carboxamides from available methyl pheophorbide *a*,<sup>#</sup> which passed without cleavage of exocycle

*E*. The combination of this approach with the known method of nucleophilic opening of exocycle *E*<sup>[9]</sup> by amines allows easily prepare regioisomeric 13<sup>1</sup>- and 15<sup>2</sup>-carboxamides of chlorin *e*<sub>6</sub> comprising various substituents in amide moieties (Scheme 1).

We discovered that interaction of methyl pheophorbide *a* **1** with 5 equivalents of amine (benzyl amine and hexadecyl amine were used) in boiling dioxane for 8 h led to 15<sup>2</sup>-carboxamides **2** and **3**, respectively.\* The presence of exocyclic ring in compounds **2** and **3** was unequivocally demonstrated by absorption spectra (Figure 1), as well as by <sup>13</sup>C NMR spectra, displayed resonances characteristic for carbonyl groups (192.06 ppm and 192.50 ppm for compounds **2** and **3**, respectively). The yield of target products **2** and **3** in this reaction exceeds 80 %; besides we have found chlorin derivatives comprising opened exocyclic ring (8-11 %). These by-products were identified as 13<sup>1</sup>-carboxamides



Scheme 1.



**Figure 1.** Absorption spectra of compounds **3** and **5** in  $\text{CH}_2\text{Cl}_2$  (spectra of compounds **2** and **4** were identical to those for related counterparts).

**4** and **5\*** (identification was carried out by comparison of their HRMS,  $^1\text{H}$  NMR and absorption spectra with those for authentic samples **4** and **5** prepared according to reported methods [20-22]).

We concluded that nucleophilic substitution of 15<sup>2</sup>-methyl ester for amine and nucleophilic opening of exocyclic ring by amine in methyl pheophorbide *a* **1** are independent. The use of non volatile amines such as benzyl amine and hexadecyl amine, high temperature of reaction, and effect of  $\beta$ -carbonyl group in methyl pheophorbide *a* **1** promote formation of 15<sup>2</sup>-carboxamide, which proceeds without exocyclic ring cleavage. On the other hand, prolong reaction at a relatively low temperature (THF, 35-40°C, 14-72 h, depending on the structure of amine used) is in favor to nucleophilic opening of exocyclic ring.

Exocyclic ring in 15<sup>2</sup>-carboxamides **2** and **3** was more resistant to nucleophilic opening by amines in comparison with that in methyl pheophorbide *a* **1**. Independent experiment showed that incubation of 15<sup>2</sup>-carboxamide **3** with benzyl amine in THF at 40°C for 72 h resulted in only traces of related 13<sup>1</sup>,15<sup>2</sup>-dicarboxamide (< 3%). This difference in reactivity of exocycle ring *E* in compounds **1** and **3** towards amines was apparently due to different electron withdrawing effects of either ester, or amide groups.

All indicated compounds **2-5** were isolated in pure forms, their structures were fully confirmed by HRMS,  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR and absorption spectra. Taken together, data presented above revealed that reaction of methyl pheophorbide *a* **1** with amines allows simple preparation of either 13<sup>1</sup>-carboxamides with opened exocycle *E*, or 15<sup>2</sup>-carboxamides with saved exocycle *E*, depending on reaction conditions. The choice of conditions is very important: recent publications [22,23] revealed that reaction of methyl pheophorbide *a* **1** with secondary amines in boiling toluene for 1.5-2 h led to mixtures of related 13<sup>1</sup>-carboxamides with opened exocycle *E* and 15<sup>2</sup>-carboxamides with saved exocycle *E*.

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## Notes and References

# Absorption spectra were registered with a ‘Thermospectronic Helios  $\alpha'$ ’ spectrophotometer in  $\text{CH}_2\text{Cl}_2$ ;  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectra - with an ‘AMX-III’ 400 MHz Bruker instrument in  $\text{CDCl}_3$ ; high resolution mass spectra (HMRS) – with a Bruker ‘Apex Ultra’ FT ICR MS instrument at ion positive electro spray ionization mode. Flash chromatography was performed on silica gel G (0.015-0.040 mm), analytical TLC – on UV254-HPTLC silica gel plates, preparative TLC – on UV254-PTLC silica gel plates “Merck”; methyl pheophorbide *a* **1** was isolated from *Spirulina platensis*.

\* Methyl 15<sup>2</sup>-(benzylcarbamoyl)pheophorbide *a*, **2**. HRMS, calculated for  $[\text{C}_{42}\text{H}_{44}\text{N}_5\text{O}_4]^+$ : 682.3393, found: 682.3378.  $^1\text{H}$  NMR  $\delta_{\text{H}}$  ppm: 8.56, 9.38, 9.51 (each 1H, s, 5-, 10-, 20-H), 7.99 (1H, dd,  $J = 11.6$  Hz and  $J = 17.8$  Hz, 3<sup>1</sup>-H), 7.42 (2H, d,  $J = 7.4$  Hz, *o*-Ph), 7.33 (2H, t,  $J = 7.4$  Hz, *m*-Ph), 7.26 (1H, t,  $J = 7.4$  Hz, *p*-Ph), 6.28 (1H, dd,  $J = 17.8$  Hz and  $J = 1.5$  Hz, 3<sup>2</sup>-H, *cis*), 6.17 (1H, dd,  $J = 11.6$  Hz,  $J = 1.5$  Hz, 3<sup>2</sup>-H, *trans*), 4.67 (2H, m,  $\text{CH}_2\text{Ph}$ ), 4.45 (2H, dt,  $J = 8.7$  Hz and  $J = 2.0$  Hz, 17<sup>1</sup>-H), 4.38 (1H, qd,  $J = 7.2$  Hz,  $J = 2.0$  Hz, 8<sup>1</sup>-H), 3.23, 3.40, 3.55, 3.68 (each 3H, s, 2-, 7-, 12- and O- $\text{CH}_3$ ), 2.20-2.34 (2H, m, 17<sup>2</sup>-H), 1.81 (3H, d,  $J = 7.3$  Hz, 18- $\text{CH}_3$ ); 1.70 (3H, t,  $J = 7.2$  Hz, 8<sup>2</sup>- $\text{CH}_3$ ), -1.60 (1H, s, NH).  $^{13}\text{C}$  NMR  $\delta_{\text{C}}$  ppm: 11.25; 12.09; 12.14; 17.40; 19.48; 23.07; 29.97; 30.92; 44.19; 50.25; 51.20; 51.68; 65.44; 93.24; 97.47; 104.29; 105.31; 122.71; 127.41; 127.96; 128.58; 128.69; 128.92; 129.16; 136.13; 136.27; 136.55; 137.95; 138.47; 138.75; 141.98; 145.19; 149.93; 150.95; 155.64; 162.73; 167.52; 172.26; 173.74; 192.06. UV-vis  $\lambda_{\text{max}}$  nm ( $\epsilon$ ): 411 (96300); 504 (8200); 535 (7000); 604 (5200); 668 (35300).

Methyl 15<sup>2</sup>-(hexadecylcarbamoyl)pheophorbide *a*, **3**. HRMS, calculated for  $[\text{C}_{51}\text{H}_{70}\text{N}_5\text{O}_4]^+$ : 816.5428, found: 816.5445.  $^1\text{H}$  NMR  $\delta_{\text{H}}$  ppm: 8.56, 9.38, 9.50 (each 1H, s, 5-, 10-, 20-H), 7.99 (1H, dd,  $J = 11.6$  Hz and  $J = 17.8$  Hz, 3<sup>1</sup>-H); 6.95 (1H, t,  $J =$

7.0 Hz, NH); 6.28 (1H, dd,  $J = 17.8$  Hz and  $J = 1.5$  Hz, 3<sup>2</sup>-H, *cis*); 6.16 (1H, dd,  $J = 11.6$  Hz,  $J = 1.5$  Hz, 3<sup>2</sup>-H, *trans*); 4.55 (2H, dt,  $J = 8.7$  Hz and  $J = 2.0$  Hz, 17<sup>1</sup>-H); 4.40 (1H, qd,  $J = 7.2$  Hz,  $J = 2.0$  Hz, 8<sup>1</sup>-H); 3.23, 3.39, 3.56, 3.67 (each 3H, s, 2-, 7-, 12- and O-CH<sub>3</sub>); 2.21-2.34 (2H, m, 17<sup>2</sup>-H); 1.88 (3H, d,  $J = 7.3$  Hz, 18-CH<sub>3</sub>); 1.70 (3H, t,  $J = 7.2$  Hz, 8<sup>2</sup>-CH<sub>3</sub>); 1.25 (br., CH<sub>2</sub> in hexadecyl moiety); 0.86 (3H, as. t, CH<sub>3</sub> in hexadecyl moiety); -1.60 (br. s, NH). <sup>13</sup>C NMR δ<sub>c</sub> ppm: 11.32; 12.16; 14.17; 17.47; 19.58; 22.77; 23.21; 23.80; 29.44; 29.64; 29.69; 29.74; 29.77; 29.85; 30.12; 31.05; 32.01; 37.14; 38.70; 40.38; 50.33; 51.32; 51.71; 53.48; 65.41; 67.92; 93.33; 97.52, 104.32; 105.56; 122.76; 128.70; 128.94; 129.28; 130.95; 131.92; 136.18; 136.22; 136.34; 136.66; 138.03; 142.05; 145.26; 149.98; 151.02; 155.72; 163.08; 167.35; 172.37; 173.80; 192.50. UV-vis λ<sub>max</sub> nm (ε): 411 (96300); 506 (8400); 539 (7300); 611 (5700); 669 (35900).

★ *13<sup>1</sup>-(Benzylcarbamoyl)chlorin*, **4**. HRMS, calculated for [C<sub>43</sub>H<sub>48</sub>N<sub>5</sub>O<sub>5</sub>]<sup>+</sup>: 714.8845; found: 714.8852. <sup>1</sup>H NMR δ<sub>H</sub> ppm: 8.80, 9.61, 9.86 (each 1H, s, 5-, 10-, 20-H); 8.05 (1H, dd,  $J = 11.6$  Hz and  $J = 17.8$  Hz, 3<sup>1</sup>-H); 7.55 (2H, d,  $J = 7.4$  Hz, *o*-Ph); 7.42 (2H, t,  $J = 7.4$  Hz, *m*-Ph); 7.34 (1H, t,  $J = 7.4$  Hz, *p*-Ph); 6.81 (1H, t,  $J = 7.0$  Hz, NH); 6.33 (1H, dd,  $J = 17.8$  Hz and  $J = 1.5$  Hz, 3<sup>2</sup>-H, *cis*); 6.12 (1H, dd,  $J = 11.6$  Hz,  $J = 1.5$  Hz, 3<sup>2</sup>-H, *trans*); 5.28 and 5.55 (each 1H, d,  $J = 18.9$  Hz, 15<sup>1</sup>-H); 4.78 (1H, dd,  $J = 5.0$  Hz and  $J = 14.5$  Hz, 17<sup>2</sup>-H); 5.06 (1H, dd,  $J = 6.1$  Hz and  $J = 14.5$  Hz, 17<sup>2</sup>-H); 4.47 (2H, m, CH<sub>2</sub>Ph); 4.39 (1H, qd,  $J = 7.2$  Hz,  $J = 2.0$  Hz, 8<sup>1</sup>-H); 3.29, 3.48, 3.53, 3.62, 3.72 (each 3H, s, 2-, 7-, 12- and O-CH<sub>3</sub>); 2.22-2.35 (2H, m, 17<sup>2</sup>-H); 1.72 (3H, d,  $J = 7.3$  Hz, 18-CH<sub>3</sub>); 1.71 (3H, t,  $J = 7.2$  Hz, 8<sup>2</sup>-CH<sub>3</sub>); -1.77 (s, NH). <sup>13</sup>C NMR δ<sub>c</sub> ppm: 11.35; 12.03; 12.18; 17.72; 19.72; 23.09; 23.91; 29.15; 29.82; 31.24; 38.03; 44.93; 49.34; 51.63; 52.19; 53.19; 67.66; 93.71; 98.90; 101.46; 107.94; 121.64; 127.84; 128.36; 128.96; 129.53; 129.96; 130.24; 134.64; 134.93; 135.03; 135.08; 136.13; 137.99; 138.99; 144.82; 149.16; 154.33; 166.72; 168.91; 169.37; 173.55; 174.02. UV-vis λ<sub>max</sub> nm (ε): 402 (104000); 500 (9100); 528 (2000); 607 (3000); 663 (35800).

*13<sup>1</sup>-(Hexadecylcarbamoyl)chlorin*, **5**. HRMS, calculated for [C<sub>52</sub>H<sub>74</sub>N<sub>5</sub>O<sub>5</sub>]<sup>+</sup>: 848.5690; found: 848.5673. <sup>1</sup>H NMR δ<sub>H</sub> ppm: 8.80, 9.63, 9.69 (each 1H, s, 5-, 10-, 20-H); 8.08 (1H, dd,  $J = 11.6$  Hz and  $J = 17.8$  Hz, 3<sup>1</sup>-H); 6.40 (1H, t,  $J = 7.0$  Hz, NH); 6.35 (1H, dd,  $J = 17.8$  Hz and  $J = 1.5$  Hz, 3<sup>2</sup>-H, *cis*); 6.13 (1H, dd,  $J = 11.6$  Hz,  $J = 1.5$  Hz, 3<sup>2</sup>-H, *trans*); 4.55 (dt,  $J = 8.7$  Hz and  $J = 2.0$  Hz, 17<sup>1</sup>-H); 4.40 (1H, qd,  $J = 7.2$  Hz,  $J = 2.0$  Hz, 8<sup>1</sup>-H); 3.31, 3.48, 3.56, 3.60, 3.80 (each 3H, s, 2-, 7-, 12- and O-CH<sub>3</sub>); 2.21-2.34 (2H, m, 17<sup>2</sup>-H); 1.72 (3H, d,  $J = 7.3$  Hz, 18-CH<sub>3</sub>); 1.74 (3H, t,  $J = 7.2$  Hz, 8<sup>2</sup>-CH<sub>3</sub>); 1.25 (br., CH<sub>2</sub> in hexadecyl moiety); 0.88 (3H, as. t, CH<sub>3</sub> in hexadecyl moiety); -1.79 (br. s, NH). <sup>13</sup>C NMR δ<sub>c</sub> ppm: 11.42; 12.08; 12.22; 14.19; 17.68; 19.80; 22.78; 23.16; 23.90; 27.34; 29.21; 29.46; 29.61; 29.69; 29.72; 29.76; 29.79; 29.85; 30.12; 31.24; 32.02; 32.90; 37.15; 37.99; 40.85; 48.14; 49.42; 50.10; 51.65; 52.20; 53.30; 53.48; 63.12; 93.12; 94.14; 98.90; 101.38; 102.67; 121.94; 128.84; 130.10; 134.93; 135.35; 135.93; 139.28; 144.53; 149.20; 153.50; 167.32; 169.29; 173.56; 174.11. UV-vis λ<sub>max</sub> nm (ε): 402 (104000); 500 (9100); 528 (2000); 607 (3000); 663 (35800).

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